

Antimalarial and mosquito repellent plants: insights from Burundi

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Research

Abstract

Background: The present ethnobotanical study was conducted to identify plant species used by Burundians to treat malaria and to repel mosquitoes, to compare this with existing literature, identify species which could be further investigated and discuss potential future promotion or cultivation.

Methods: Surveys were conducted between April and October 2018 in seven provinces representing the five ecological zones of Burundi. A semi-structured questionnaire was administered to 341 randomly selected respondents (between 25 and 50 household heads in each province).

Results: A total of 44 plant species were reported in this study: 32 as antimalarial, two as mosquito repellents and 10 for both purposes. For antimalarial plants (84%) and mosquito repellent plants (88%), leaves were the most commonly used plant part. According to the respondents, 28 plant species were being cultivated and 16 were mostly collected from the wild. An examination of the literature on some of the plant species mentioned in this study revealed that eight of them had never been studied before.

Conclusions: The use of antimalarial and mosquito repellent plants in Burundi was highlighted in this study. Its goal is to create a database of antimalarial and mosquito repellent plants. This will aid decision-making in the development of traditional medicine and the conservation of medicinal plants.

Keywords: Ethnobotany; antimalarial activity; mosquito repellents; plants cultivation; Eco-climatic zones.

Background

Malaria is the world's deadliest disease with, approximately 229 million malaria cases and 409 thousand deaths reported worldwide in 2019, with the African region accounting for 94% of these deaths (WHO 2020). Burundi is

one of the countries most affected by this disease, which has been identified as the country's leading cause of morbidity and mortality (République du Burundi 2018). In 2017, for example, the annual cumulative number of malaria deaths was 815 per 1,000 (Sinzinkayo 2018). Surprisingly, the treatment of malaria is still not subsidized in Burundi. Ndayizeye *et al.* (2020) reported that the low-income population (e.g. Twa hunter-gather community of Pygmy origin) considers access to western medicine to be limited due to its prohibitively high cost, despite the fact that malaria treatment is subsidized in some countries (e.g. Kenya) (Delbanco *et al.* 2017). In this context, the use of medicinal plants in the treatment of malaria becomes an important contribution to the health of people who cannot afford the pharmaceutical products from conventional medicine.

Natural plant recipes are commonly used to treat malaria in Africa (for example, Ghana and Burkina Faso) (Ankrah *et al.* 2003; Bonkian *et al.* 2017). Furthermore, recent research indicates that the malaria parasite and its vector are becoming increasingly resistant to conventional drug molecules developed by industrialists (Lutgen *et al.* 2018; WHO 2018). Nonetheless, new tools and products for the detection, treatment and prevention of malaria are constantly being developed (Hemingway *et al.* 2016; Othman *et al.* 2018). As a result, research on malaria alternatives remains current and is expected to mobilize scientists in a variety of fields. Several studies have been conducted to determine which plant species are used as antimalarials. For example, in northeast India, (Namsa *et al.* 2011) investigated anti-malarial herbal remedies and (Odoh *et al.* 2018) identified medicinal plants used by the people of Nsukka to the treat malaria in south-eastern Nigeria.

Furthermore, because prevention is better than cure, the role of plants in avoiding mosquito's bites has been investigated. Some studies have focused on malaria vector control using plants, for example. in the endemic regions of Cameroon (Youmsi *et al.* 2017), the north-eastern Tanzania (Kweka *et al.* 2008), in Dai people of Xishuangbanna in China (Gou *et al.* 2020) and the uMkhanyakude district, KwaZulu-Natal in South Africa (Mavundza *et al.* 2011).

Two recent studies from Burundi focused on the characteristics of traditional healers (Falisse *et al.* 2018) and traditional healers' use of plants to treat microbial diseases (Ngezahayo *et al.* 2015). Malaria was overlooked in both studies. In general, traditional healers are not visited to treat malaria because it is a common disease and people in Burundi, particularly the poor, prefer to self-medicate. Apart from understanding which species is used, it is also important to understand (i) plant part utilized, (ii) habitat and (iii) perceived abundance in the wild (if not cultivated). These three aspects can work in favor of actions to manage and conserve species and ecosystems. In fact, harvesting the wood, bark or entire medicinal plant is more destructive and may result in the death of the individual plant, whereas the fruits and the leaves pose less of a threat to the species (Delbanco *et al.* 2017). Similarly, harvesting habitat (for example, the forest versus fallow) of medicinal plants influences the potential threat on the ecosystem. The perception of abundance or scarcity is relevant for conservation policy-making and/or may inspire seedling production and population dissemination programs.

This study focused on plant species used by Burundians to treat malaria and repel mosquitoes. The objectives of this study are: (i) to identify plant species used by Burundians to treat malaria and repel mosquitoes, (ii) to compare this with the literature to identify species whose usage has not been reported and which could be further investigated, (iii) to identify part (s) used, habitat, and perceived abundance in the wild to discuss potential future promotion/cultivation of some species.

Material and Methods

Study area

Burundi has a population of nearly 12 million people and a population density of 422 inhabitants per km² (PopulationData.net - Burundi). The altitude ranges from 774 m on the shores of Lake Tanganyika to 2680 m on the mountain ranges, gradually decreasing to 1100 m in the east of the country (Nzigidahera 2012).

This study was conducted in seven of the 18 provinces of Burundi (Fig. 1). The selected provinces represent the five eco-climatic zones of Burundi as described by Martens and Sauttiaux (1979) in (Nzigidahera 2012): (i) the Imbo plain, with elevations ranging from 800 to 1100 m and mean annual temperature exceeding 23 °C, (ii) the Mirwa foothills, with elevations ranging from 1000 to 1700 m and mean annual temperature ranging from 18 to 28 °C, (iii) Congo-Nile ridge with elevations ranging from 1700 to 2684 m and mean annual temperatures ranging from 14 to 15 °C, (iv) the central highlands with elevations ranging from 1350 to 2000 m and mean annual temperature ranging from 17 to 20 °C; and (v) the northern and eastern lowlands with elevations ranging from 1100 to 1400 m and mean annual temperatures ranging from 20 to 23 °C. The Imbo plain and Mirwa foothills are represented by Bubanza, Bujumbura Mairie and Rumonge while in the Congo-Nile ridge, Bubanza and Muramvya were selected. The Central

highlands was represented by Muramvya, Ngozi and Cankuzo; and the Northern and eastern lowlands were represented by Cankuzo and Kirundo. Burundi's population is made up of three major ethnic groups: Twa (hunter-gatherers of Pygmy origin), Tutsi, and Hutu (both farmers of Bantu origin). We only interviewed the last two groups in this study.

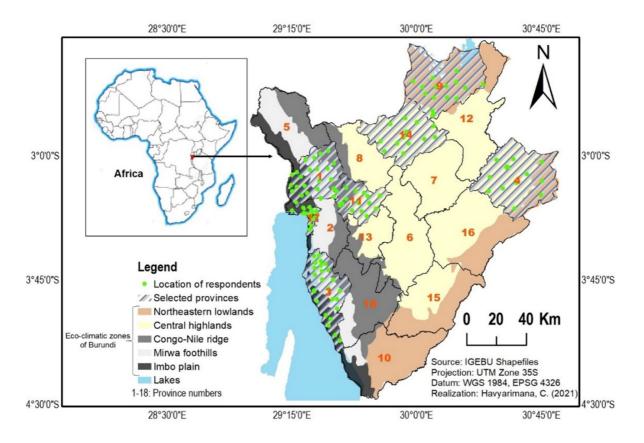


Figure 1. Eco-climatic zones of Burundi and the selected provinces: Bubanza (1), Rumonge (3), Cankuzo (4), Kirundo (9), Muramvya (11), Ngozi (14) and Bujumbura municipality (17).

Ethnobotanical data collection

Semi-structured questionnaires were administered to 341 respondents between April and October 2018. We targeted one village (rural) and one urban center in each province, and we randomly selected between 25 and 50 household heads (aged above 18 years) who admitted to using at least one plant as natural mosquito repellent or as antimalarial.

Respondents were first informed that the aim of the study was to better understand the importance of natural mosquito repellent and antimalarial plants, and that their responses would be kept anonymous and confidential. The questionnaire was given to them if they agreed to participate in the study. The semi-structured questionnaire addressed: (i) respondents' socioeconomic characteristics (e.g. gender, age, urban or rural residence); (ii) species' used, plant part, and type of usage; and (iii) the ecology and availability of the species mentioned. The first author facilitated all questionnaires, which were done in Kirundi. Smap Server Version 19.04 installed in an android phone was used to collect data before being exported to Microsoft Excel 2016 software. To conduct this research, we obtained a research permit and permission from local authorities in each village. We informed all informants of our intentions and obtained their verbal consent.

Plant samples were collected around the villages and deposited at the Herbarium of the University of Burundi (BJA) for identification and authentication by a plant taxonomist. Plant species' scientific names mentioned during the interviews were verified from the collected plant specimens. Several resources (Troupin 1982; Reekmans and Niyongere 1983; Troupin 1983; Troupin 1985; Troupin 1988; Lebrun and Stork 1991) were used for plant identification. Voucher specimen numbers for 35 out of the 44 species listed in this study are indicated in Table 1. The nine species with no herbarium samples are commonly cultivated plants in Burundi.

Ethnobotanical indices

The relative importance of the species cited was expressed as the percentage of respondents who cited that species as following: $F = \frac{S}{N} * 100$ (Ndiaye *et al.* 2017) with F: species citation frequency; S: number of species citations and N: total number of interviewees.

Finally, based on the plant species here inventoried, a literature review was conducted to highlight the overview knowledge of their anti-malaria and anti-mosquito's properties.

In order to determine if respondents were using plant species of conservation concern, the conservation status of all the species mentioned in the study was checked using the IUCN Red List (IUCN 2019). A literature review was also conducted to check the mosquito repellent or antimalarial uses, ecology and distribution of the species mentioned by study participants. Plant nomenclature follows the African Plant Database (version 3.4.0, African Plant Database, 2020).

Statistical analysis

We investigated if there were differences in the number of species cited across genders (male or female), age groups (<35 years, 35 to 59 years, > 60 years) or domicile (urban or rural). We used Chi-square (χ^2) test, with significance level α =0.05. Overall, we interviewed 136 males and 205 females, 132 urban residents and 209 rural residents and 102, 127 and 112 respondents of <35 years, 35 to 59 years, > 60 years respectively.

Results and Discussion

Species cited

Respondents identified 44 plant species in total, 32 of which were used as antimalarial, two as mosquito repellents and ten for both purposes (Table 1). Other studies reported 107 species used as antimalarial in Uganda (Okello and Kang 2019), 35 in Rwanda (Muganga *et al.* 2010) and 139 in Kenya (Omara 2020). Most species were also mentioned by participants in other studies in 12 other tropical African countries, namely: Tanzania in the Kagera and Lindi regions (Nondo *et al.* 2015), Uganda around the Mabira Forest Reserve (Tugume *et al.* 2016), Benin in the Allada Plateau (Yetein *et al.* 2013), Nigeria in the States of Ondo, Ogbomoso and Ogun (Idowu *et al.* 2010; Olorunnisola *et al.* 2013; Oyeyemi *et al.* 2019), Namibia in the Oshikoto region (Cheikhyoussef *et al.* 2011), the Democratic Republic of Congo in the Mbanza-Ngungu region (Nzuki 2016), Ethiopia in the districts of Hawassa Zuria and Shinile (Mesfin *et al.* 2012; Tefera and Kim 2019), Mali in the district of Bamako (Dénou *et al.* 2017), Burkina Faso in the Sahel region (Bonkian *et al.* 2017), Togo in the maritime region (Koudouvo *et al.* 2011), Morocco in the Talassemtane National Park (Rhattas *et al.* 2016) and Ivory Coast in the Zanzan District (Kouadio *et al.* 2016).

The most commonly cited antimalarial species were: *Gymnanthemum amygdalinum* (Delile) Sch. Bip. ex Walp (F=57.48), *Eucalyptus globulus* subsp. *maidenii* (F. Muell.) Kirkp. (F=14.96), *Cinchona officinalis* L. (F=10.56), *Cajanus cajan* (L.) Huth (F=9.38) and *Tetradenia urticifolia* (Hochst.) Codd (F=9.09).

Cymbopogon citratus (DC.) Stapf (F=6.16) was the most frequently mentioned mosquito repellent species. Mosquito repellent species were mentioned by fewer people than antimalarial species. Thirty-six active compounds were also reported to have antimalarial or mosquito repellent properties. These excluded eight species: *Markhamia lutea* (Benth.) K. Schum., *Chenopodium ugandae* (Aellen) Aellen, *Euphorbia grantii* Oliv., *Plectranthus esculentus* N.E.Br., *Eucalyptus globulus* subsp. *maidenii* (F. Muell.), *Cencrhus purpureum* (Schumach.) Morrone, *Digitaria abyssinica* (Hochst. ex A.Rich.) Stapf and *Solanum terminale* Forssk, for which we recommend further work to validate their usages.

Male respondents identified 44 species, while women identified 40. Urban respondents identified 39 species, while rural respondents identified 42. In terms of age groups, 112 respondents of <35 years, 35 to 59 years, > 60 years named 31, 39, and 44 species, respectively. The calculated Khi-deux value is less than the critical Khi-deux value in terms of age and residence. However, when gender is considered, the calculated Khi-deux value is greater than the critical Khi-deux value (Table 2).

In this study, gender and residence (urban/rural) had no significant effects on the knowledge of anti-malarial plants. A study from autonomous district of Abidjan (Ivory Coast) showed that women identified more medicinal plants than men (Manouan *et al.* 2014), another study from the Edough peninsula (north-east Algeria) also showed that women hold greater traditional phytotherapeutic knowledge than men (Hamel *et al.* 2018). A study from Quebradas del Norte (Uruguay) showed that rural residents identified more medicinal plants than urban residents, but the differences were not statistically significant (Latorre *et al.* 2018).

Table 1. Antimalarial and mosquito repellent plants used in Burundi: Taxonomy position, local name, frequency of citation

| Family | Plant species | Local name | Voucher specimen number | F (%) Respondents anti-malaria | F (%) Respondents mosquito repellent | Mentioned by respondents elsewhere | Anti-malaria or mosquito repellent property reported in literature |
|---------------|---|--------------------------------------|------------------------------------|--------------------------------------|---|--|--|
| Anacardiaceae | <i>Mangifera indica</i> L. | Umwembe | - | 2,64 | 0 | (Nondo <i>et al.</i> 2015; Yetein <i>et al.</i> 2013; Oyeyemi <i>et al.</i> 2019) | (Cudjoe <i>et al.</i> 2020) |
| Asphodelaceae | Aloe sp. | lgikakarubamba, Impfizi y'umusozi | - | 2,35 | 0 | (Teka <i>et al.</i> 2016; Amir <i>et al.</i> 2019; Mesfin <i>et al.</i> 2012; Kasali 2014; Dibessa et al. 2020; Geremedhin, Bisrat, and Asres 2014; Michayewicz 2013) | (Geremedhin, Bisrat, and Asres 2014; Dibessa <i>et al.</i> 2020; Teka <i>et al.</i> 2016) |
| Asteraceae | <i>Artemisia annua</i> L. | Aritemiziya | - | 1,17 | 0 | (Manya et al. 2020; Feng <i>et al.</i> 2020; Chiribagula <i>et al.</i> 2020) | (Alesaeidi and Miraj 2016; Czechowski <i>et al.</i> 2019; Elfawal <i>et al.</i> 2012; Liu <i>et al.</i> 1992) |
| Asteraceae | <i>Baccharoides lasiopus</i> (O. Hoffm.) H. Rob | Umuvuma | HC&NK023 | 1,76 | 0 | (Amuka <i>et al.</i> 2014; Mulei, Otieno, and Onkware 2014; Maroyi 2020) | (Muregi <i>et al.</i> 2007) |
| Asteraceae | <i>Bidens pilosa</i> L. | Icanda | HC&NK040 | 2,05 | 0,29 | (Clement <i>et al.</i> 2020) | (Nadia <i>et al.</i> 2020) |
| Asteraceae | <i>Erigeron sumatrensis</i> (S.F. Blake) Pruski & G.Sancho | Umururasase | HC&NK027 | 4,69 | 0 | (Manya <i>et al.</i> 2020) | (Boniface <i>et al.</i> 2015) |
| Asteraceae | <i>Guizotia scabra</i> (Vis.) Chiov. | Ikizimyamuriro | HC&NK031 | 0,88 | 0 | (Nondo <i>et al.</i> 2015) | (Nondo <i>et al.</i> 2015) |
| Asteraceae | <i>Gutenbergia cordifolia</i> Benth. ex Oliv. | Umweza | HC&NK004 | 1,47 | 0 | (Koch <i>et al.</i> 2005) | (Koch <i>et al.</i> 2005) |
| Asteraceae | <i>Gymnanthemum amygdalinum</i> (Delile) Sch. Bip. ex Walp | Umubirizi, Umufumya | HC&NK009, HC&NK010, HC&NK011 | 57,48 | 0 | (Nondo <i>et al.</i> 2015; Yetein <i>et al.</i> 2013; Oladeji <i>et al.</i> 2020) | (Bihonegn <i>et al.</i> 2019) |

| Family | Plant species | Local name | Voucher specimen number | F (%) Respondents anti-malaria | F (%) Respondents mosquito repellent | Mentioned by respondents elsewhere | Anti-malaria or mosquito repellent property reported in literature |
|----------------|---|---|------------------------------------|--------------------------------------|---|---|---|
| Asteraceae | <i>Solanecio mannii</i> (Hook.f.) C.Jeffrey | Umutagari, Umugango | HC&NK017, HC&NK018, HC&NK019 | 4,11 | 0 | (Manya <i>et al.</i> 2020) | (Muganga <i>et al.</i> 2010) |
| Asteraceae | <i>Tithonia diversifolia</i> (Hemsl.) A. Gray | lkinyamuhora, Banyakuyumye, Kivyeyi | HC&NK026 | 7,91 | 0,29 | (Odugbemi <i>et al.</i> 2011) | (Afolayan, Oladokun, and Fasoranti 2020) |
| Bignoniaceae | <i>Markhamia lutea</i> (Benth.) K. Schum. | Umusave | HC&NK001, HC&NK002, HC&NK003 | 2,64 | 0 | (Nondo <i>et al.</i> 2015; Tugume <i>et al.</i> 2016) | - |
| Caricaceae | <i>Carica papaya</i> L. | lgipapayi | - | 1,17 | 0 | (Olorunnisola <i>et al.</i> 2013; Dike, Obembe, and Adebiyi 2012) | (Abdillah <i>et al.</i> 2015; Priyadarshi and Ram 2018) |
| Casuarinaceae | <i>Casuarina equisetifolia</i> L. | Akajwari | HC&NK032 | 0 | 0,88 | - | (Malann <i>et al.</i> 2015) |
| Chenopodiaceae | <i>Chenopodium ambrosioides</i> L. | Akavunjahoma, Umusuzi w'ingona | NK&BU011 | 3,82 | 0 | (Yetein <i>et al</i> . 2013; Frausin <i>et al</i> . 2015) | (Cysne <i>et al.</i> 2016) |
| Chenopodiaceae | <i>Chenopodium ugandae</i> (Aellen) Aellen | Umugombe | HC&NK033 | 1,47 | 0 | (Chiribagula <i>et al.</i> 2020) | - |
| Cupressaceae | Cupressus sp. | Isederi | HC&NK016 | 0 | 2,64 | (Kasali 2014) | (Saad <i>et al</i> . 2017) |
| Euphorbiaceae | <i>Euphorbia grantii</i> Oliv. | Imambura | HC&NK015 | 0,29 | 0,88 | - | - |
| Fabaceae | <i>Cajanus cajan</i> (L.) Huth | Intengwa, Umukunde, Incaruzo | HC&NK022 | 9,38 | 0 | (ldowu <i>et al.</i> 2010; Oyeyemi <i>et al.</i> 2019) | (Ajaiyeoba <i>et al.</i> 2013; Olusi, Ibukunoluwa, and Dada 2016) |
| Fabacea | <i>Senna didymobotrya</i> (Fresen.) H. S. Irwin & Barneby | Umubagabaga | HC&NK013 | 4,11 | 0 | (Njoroge and Bussmann 2006) | (Waiganjo <i>et al</i> . 2020) |
| Fabaceae | <i>Sesbania sesban</i> (L.) Merr. | Umunyegenyege | HC&NK005 | 2,93 | 0 | (Shah and Rahim 2017) | (El-Emam, Mahmoud, and Bayaumy 2015) |
| Lamiaceae | <i>Ocimum gratissimum</i> L. var. <i>gratissimum</i> | Kabugagwe, Simama nikwambiye | HC&NK007, HC&NK008 | 1,73 | 1,76 | (Oyeyemi <i>et al.</i> 2019; Oladeji <i>et al.</i> 2020; | (Zirihi <i>et al.</i> 2005) |

| Family | Plant species | Local name | Voucher specimen number | F (%) Respondents anti-malaria | F (%) Respondents mosquito repellent | Mentioned by respondents elsewhere | Anti-malaria or mosquito repellent property reported in literature |
|----------------|---|------------------|-------------------------------|--------------------------------------|---|--|---|
| | | | | | | Odugbemi <i>et al.</i> 2011) | |
| Lamiaceae | Plectranthus barbatus | lgicuncu | HC&NK035, HC&NK036 | 2,05 | 0 | (Kiraithe <i>et al.</i> 2016; Philip <i>et al.</i> 2017) | (Owuor <i>et al</i> . 2012) |
| Lamiaceae | <i>Plectranthus esculentus</i> N.E.Br. | Inumpu | - | 1,76 | 0 | (Frausin <i>et al</i> . 2015) | - |
| Lamiaceae | <i>Tetradenia urticifolia</i> (Hochst.) Codd | Umuravumba | HC&NK006 | 9,09 | 0,29 | (Bahekar and Kale 2013) | (Bickii <i>et al.</i> 2007; Noronha <i>et al.</i> 2020) |
| Lauraceae | <i>Persea americana</i> Mill. | lvoka | - | 1,76 | 0 | (Philip <i>et al.</i> 2017; Idowu <i>et al.</i> 2010) | (Adesina <i>et al.</i> 2016) |
| Meliaceae | <i>Azadirachta indica</i> A. Juss. | Arobayine | HC&NK039 | 5,57 | 0,59 | (Dike, Obembe, and Adebiyi 2012; Noronha <i>et al.</i> 2020; Bodeker <i>et al.</i> 2001; Iyamah and Idu 2015) | (Akpuaka <i>et al</i> . 2013; Deshpande, Gothalwal, and Pathak 2014; Murugan <i>et al.</i> 2016) |
| Moringaceae | <i>Moringa oleifera</i> Lam. | Moringa | - | 1,76 | 0 | (Oladeji <i>et al.</i> 2020; Nondo <i>et al</i> . 2015) | (Cudjoe <i>et al.</i> 2020) |
| Myrtaceae | <i>Eucalyptus globulus</i> subsp. <i>Maidenii</i> (F. Muell.) | Umukaratusi wera | HC&NK028 | 14,96 | 2,35 | (Tefera and Kim 2019) | - |
| Myrtaceae | Psidium guajava | Ipera | HC&NK020 | 2,05 | 0 | (Odugbemi <i>et al.</i> 2011; Nondo <i>et al.</i> 2015) | (Kaushik <i>et al.</i> 2015) |
| Phytolaccaceae | <i>Phytolacca dodecandra</i> L. Hér. | Umwokora | HC&NK029, HC&NK030 | 2,93 | 0,59 | (Bahekar and Kale 2013; Gurmu <i>et al.</i> 2018) | (Adinew 2014) |
| Poaceae | <i>Cencrhus purpureum</i> (Schumach.) Morrone | Urubingo | HC&NK021 | 1,17 | 0 | (Nondo <i>et al.</i> 2015; Oladeji <i>et al.</i> 2020) | - |
| Poaceae | <i>Cymbopogon citratus</i> (DC.) Stapf | Сауісауі | - | 0,58 | 6,16 | (Nondo <i>et al.</i> 2015; Oladeji <i>et al.</i> 2020) | (Melariri <i>et al.</i> 2011) |

| Family | Plant species | Local name | Voucher specimen number | F (%) Respondents anti-malaria | F (%) Respondents mosquito repellent | Mentioned by respondents elsewhere | Anti-malaria or mosquito repellent property reported in literature |
|--------------|--|------------------------------------|-------------------------------|--------------------------------------|---|---|---|
| Poaceae | <i>Digitaria abyssinica</i> (Hochst. ex A.Rich.) Stapf | Urwiri | HC&NK043, HC&NK044 | 1,47 | 0 | - | - |
| Poaceae | <i>Zea mays</i> L. | lkigori | - | 7,04 | 0 | (Betti <i>et al.</i> 2013; Koudouvo <i>et al.</i> 2011) | (Okokon <i>et al.</i> 2017, 2019) |
| Polygalaceae | <i>Securidaca longipedunculata</i> Fresen. | Umunyagasozi | NK019 | 2,05 | 0 | (Bonkian <i>et al.</i> 2017) | (Bah <i>et al.</i> 2007; Ancolio <i>et al.</i> 2002) |
| Rubiaceae | <i>Cinchona officinalis</i> L. | Ikinini | HC&NK034 | 10,56 | 0 | (Süntar 2020; Selvam and Durai 2018; Chiribagula <i>et al.</i> 2020) | (Karle and Bhattacharjee 1999) |
| Rubiaceae | <i>Rubia cordifolia</i> L. subsp. <i>conotricha</i> (Gand.) Verdc. | Umukararambwa | HC&NK012 | 0,88 | 0 | (Dwivedi <i>et al.</i> 2020) | (Nyambati <i>et al.</i> 2013) |
| Rutaceae | <i>Citrus limon</i> (L.) Osbeck | Indimu | - | 1,76 | 0,88 | (Youmsi <i>et al.</i> 2017) | (Bonkian <i>et al.</i> 2019) |
| Sapindaceae | <i>Dodonea viscosa</i> Jacq. | Umusasa | HC&NK025 | 3,23 | 0 | (Tefera and Kim 2019) | (Clarkson <i>et al.</i> 2004) |
| Solanaceae | <i>Physalis angulata</i> L. | Intumbaswa, Amahwibiri, Agaperi | HC&NK024 | 2,35 | 0 | (Odugbemi <i>et al.</i> 2011; Frausin <i>et al.</i> 2015) | (Lusakibanza <i>et al.</i> 2010) |
| Solanaceae | <i>Solanum terminale</i> Forssk. | Umuhanurankuba | HC&NK037 | 0,88 | 0 | - | - |
| Verbenaceae | <i>Lantana camara</i> L. | Mavyi ya kuku | HC&NK038 | 1,17 | 0 | (Selvam and Durai 2018; Yetein <i>et al.</i> 2013; Nondo <i>et al.</i> 2015) | (Ved <i>et al.</i> 2018) |
| Verbenaceae | <i>Lantana trifolia</i> L. | Umuhengerihengeri | HC&NK014 | 1,17 | 0 | (Mukungu <i>et al.</i> 2016) | (Seyfe <i>et al.</i> 2017) |

| Variable | Anti-malarial | Anti-mosquito | DF | χ² (α=0,05) | χ² | P-value | Decision |
|----------------|---------------|---------------|----|-------------|--------|---------|---|
| Age | | | | | | | |
| > 35 years | 164 | 12 | 2 | 5,991 | 11,117 | 0,021 | $\chi^2 > \chi^2 (\alpha = 0.05)$ and P-value $< \alpha$; H ₀ accepted |
| 35 to 59 years | 344 | 32 | | | | | |
| > 60 years | 212 | 13 | | | | | |
| Gender | | | | | | | |
| Female | 276 | 29 | 1 | 3,841 | 1,636 | 0,079 | $\chi^2 < \chi^2 (\alpha=0,05)$ and P-value > α ; H ₀ rejected |
| Male | 444 | 28 | | | | | , |
| Residence | | | | | | | |
| Rural | 496 | 27 | 1 | 3,841 | 3,485 | 0,125 | $\chi^2 < \chi^2 (\alpha = 0,05)$ and P-value > α ; H ₀ rejected |
| Urban | 224 | 30 | | | | | |

Table 2. Significance of age, gender and residence variables on knowledge of antimalarial and mosquito repellent plants listed in Burundi

However, in this study, age had a significant impact on the knowledge of anti-malarial plants. A study from of the East Douala (Cameroon) showed that knowledge of the properties and uses of medicinal plants are generally acquired after a long experience (Latreche and Sadoudi 2017). Another study from Nyong department (Cameroon) showed that young people are mostly in school and lose interest in traditional medicine (Mpondo *et al.* 2017). Therefore, this study shows that age can be recognized as a determining factor in the knowledge of medicinal plants, which consideration may lead to reliable results in ethnobotanical studies.

Plant parts and preparations

The leaves of antimalarial plants were the most commonly used plant part (84%). Bark (7%), entire plants (5%), roots (2%) and sap (2%) were less frequently mentioned. Preparation modes of antimalarial plants included maceration (60%), decoction (37%) and infusion (3%). Leaves were also the most commonly used plant part (88%) in mosquito repellent plants. Entire plants (12%) were mentioned less frequently. Mosquito repellent plants are either burned (58%), hung in the house (25%), or even planted near residential areas (17%).

The results of this study agree with other studies that reported that leaves were the most commonly used plant part (Yetein *et al.* 2013; Kalonda *et al.* 2014; Bla *et al.* 2015; Sylla *et al.* 2018). In this study, maceration and decoction were the most used mode of preparation. According to other studies, maceration and decoction are the most frequently used modes of preparation (Yetein *et al.* 2013; Nzuki 2016; Sylla *et al.* 2018). The use of decoction may be justified by the fact that it promotes extraction and the release of volatile toxicants.

Species' habitats and relative abundance

Respondents reported that 28 antimalarial plants were cultivated (farms or planted forests) and 16 were mostly collected from the wild: 7 species were mostly found in semi-natural forests, 7 in savannahs, 14 mostly in fallows, and 11 in cultivated fields (Table 3). Seven of the species collected in the wild were identified as scarce by more than 70 % of those who used that species (Table 3). The available literature revealed that species perceived as scarce were, in fact, scarce in the wild. According to the IUCN Red list, none of the species cited in this study are of international conservation concern. The majority of plant species identified in this study were cultivated, with only a few collected in the wild. A study conducted on Batan Island (Philippines) showed that medicinal plants were cultivated, but the majority were found growing wild in fields, backyards, or forests (Abe and Ohtani 2013).

This study identified species which were scarce in some localities in terms of relative abundance. This is consistent with the study conducted in India, which showed that medicinal plants are becoming increasingly scarce as a result of overexploitation and unscientific harvesting practices (Akhilraj *et al.* 2021).

Literature review on some plant species mentioned in this study and their antimalarial or mosquito repellent compounds

Some plants species (nine out of 11) included in this chapter are among the most cited by traditional healers during the ethnobotanical survey. Others (*Artemisia annua* and *Azadirachta indica*), have not been widely cited (probably because they were introduced recently in Burundi, and therefore are not known in the country), but are recorded in literature as sources of potent antimalarials that have played a major role in the fight against malaria (Manya et al. 2020; Feng *et al.* 2020; Chiribagula *et al.* 2020; Dike, Obembe, and Adebiyi 2012; Noronha *et al.* 2020; Bodeker *et al.* 2001; Iyamah and Idu 2015). The structures of active molecules isolated from these different plants are presented in Table 4.

Artemisia annua L.

Artemisia annua L. (Asteraceae) is a mugwort of Chinese origin known as Qinghao and traditionally used in Chinese medicine against malarial fevers for a very long time. The structure of the active molecule (artemisinin, **1**) was elucidated in the 1970s (Collaboration Research Group for Qinghaosu 1977) after several years of research on antimalarials (large-scale phytochemical screening in animals infected with malaria). Known in China as *qinghaosu*, this molecule is a sesquiterpene lactone (carrying a peroxide group which is essential for its therapeutic efficacy), with an extremely effective anti-malarial effect and capable of inducing oxidative stress in *Plasmodium*, the causative agent of malaria. In particular, it makes it possible to treat strains resistant to chloroquine, with practically no side effects. Its discovery earned Youyou Tu (a Chinese scientist) the Nobel Prize in Physiology or Medicine in 2015 (Liu *et al.* 2015). Other types of molecules (non-terpene) have been isolated from this species; thus, flavonoids (**2-7**) have shown an antimalarial activity, as well as a potential synergistic effect with artemisinin *in vitro* (Cao *et al.* 2020; Gruessner *et al.* 2021, Zhou *et al.* 2021).

| Species | Part used | Habitat | Perceptions of respondents on Abundance according to the survey (% of respondents) | Literature for the plants perceived as scarce |
|---|------------|--|--|---|
| Collected from the wild | | | | |
| <i>Bidens pilosa</i> L. | Lf | Fallow, Cultivated fields | Abundant (100 %) | - |
| <i>Erigeron sumatrensis</i> (S.F. Blake) Pruski & G.Sancho | Lf | Cultivated fields, Savannahs | Abundant (100 %) | - |
| <i>Digitaria abyssinica</i> (Hochst. ex A. Rich.) Stapf | Lf | Cultivated fields, Savannahs, Fallow | Scarce (10 %), Less abundant (20 %), Abundant (75 %) | - |
| <i>Dodonea viscosa</i> Jacq. | Lf | Fallow, Cultivated fields | Scarce (70 %), Less abundant (25 %), Abundant (5 %) | (Ng <i>et al.</i> 2021) |
| <i>Guizotia scabra</i> (Vis.) Chiov. | Lf | Semi-natural forest, Cultivated fields, Fallow, Savannahs | Scarce (30 %), Less abundant (20 %), Abundant (50 %) | - |
| <i>Gutenbergia cordifolia</i> Benth. ex Oliv. | Lf | Fallow, Savannahs | Scarce (70 %), Less abundant (25 %) | (Kavana <i>et al.</i> 2019) |
| <i>Lantana camara</i> L. | Lf | Semi-natural forest, Savannahs, Fallow | Less abundant (5 %), Abundant (95 %) | - |
| <i>Lantana trifolia</i> L. | Lf, Rt, Ec | Cultivated fields, Savannahs, Fallow | Scarce (70 %), Less abundant (15 %), Abundant (15 %) | (Adomou 2005) |
| <i>Ocimum gratissimum</i> L. var. <i>gratissimum</i> | Wp | Cultivated fields, Fallow | Scarce (30 %), Less abundant (15 %), Abundant (55 %) | - |
| <i>Physalis angulata</i> L. | Lf | Semi-natural forest, Cultivated fields, Fallow | Scarce (30 %), Less abundant (15 %), Abundant (55 %) | - |
| <i>Phytolacca dodecandra</i> L. Hér. | Lf | Cultivated fields, Fallow, Natural forest | Scarce (70 %), Less abundant (25 %), Abundant (5 %) | (MINEAGRIE 2014) |
| <i>Rubia cordifolia</i> L. subsp. <i>conotricha</i> (Gand.) Verdc. | Lf | Natural forest, Fallow | Scarce (80 %), Less abundant (15 %), Abundant (5 %) | (Masharabu <i>et al.</i> 2010) |
| <i>Securidaca longipedunculata</i> Fresen. | Lf | Savannahs | Scarce (10 %), Less abundant (15 %), Abundant (75 %) | - |
| <i>Sesbania sesban</i> (L.) Merr. | Lf | Semi-natural forest, Cultivated fields, Fallow | Scarce (70 %), Less abundant (30 %) | (Orwa <i>et al.</i> 2009) |

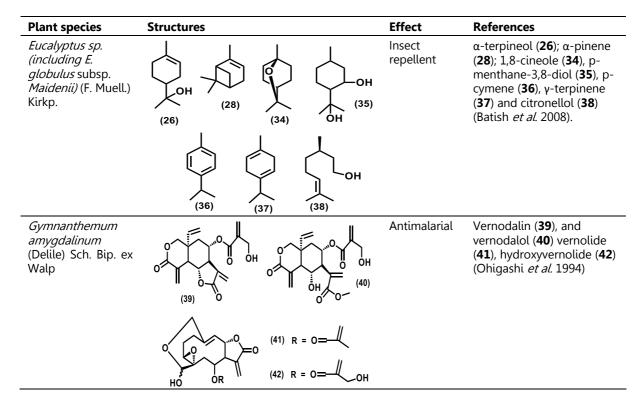
| Species | Part used | Habitat | Perceptions of respondents on Abundance according to the survey (% of respondents) | Literature for the plants perceived as scarce |
|--|-----------|---|---|--|
| <i>Solanum terminale</i> Forssk. | Lf | Semi-natural forest, Fallow | Scarce (95 %), Less abundant (5 %) | (Schmelzer and Gurib- Fakim 2008) |
| <i>Tithonia diversifolia</i> (Hemsl.) A. Gray | Lf | Fallow, Cultivated fields | Less abundant (15 %), Abundant (85 %) | - |
| Cultivated | | | | |
| Aloe sp. | Lf | Gardens, Cultivated fields | Less abundant (25 %), Almost untraceable (75 %) | - |
| <i>Artemisia annua</i> L. | Lf | Gardens, Cultivated fields | Scarce (50 %), Less abundant (25 %), Abundant (25 %) | - |
| Azadirachta indica A. Juss. | Lf, Ec | Gardens, Cultivated fields | Scarce (50 %), Less abundant (25 %), Abundant (25 %) | - |
| <i>Baccharoides lasiopus</i> (O. Hoffm.) H. Rob | Lf | Savannahs, Gardens, Cultivated fields | Scarce (60 %), Less abundant (25 %), Abundant (10 %) | - |
| <i>Cajanus cajan</i> (L.) Huth | Lf | Gardens, Cultivated fields | Scarce (60 %), Less abundant (25 %), Abundant (15 %) | - |
| <i>Carica papaya</i> L. | Lf | Gardens, Cultivated fields | Scarce (65 %), Less abundant (15 %), Abundant (20 %) | - |
| <i>Casuarina equisetifolia</i> L. | Lf | Artificial afforestation | Scarce (30 %), Less abundant (35 %), Abundant (35 %) | - |
| <i>Cencrhus purpureum</i> (Schumach.) Morrone | Lf | Little disturbed forests, Gardens, Cultivated fields | Scarce (50 %), Less abundant (25 %), Abundant (25 %) | - |
| <i>Chenopodium ambrosioides</i> L. | Wp | Cultivated fields, Gardens, Fallow | Scarce (50 %), Less abundant (25 %), Abundant (25 %) | - |
| <i>Chenopodium ugandae</i> (Aellen) Aellen | Lf | Fallow, Gardens, Cultivated fields | Scarce (70 %), Less abundant (12 %), Abundant (18 %) | - |
| <i>Cinchona officinalis</i> L. | Lf, Ec | Artificial afforestation, Cultivated fields, Gardens | Scarce (80 %), Less abundant (15 %), Abundant (5 %) | - |
| Citrus limon (L.) Osbeck | Lf | Gardens, Cultivated fields | Scarce (60 %), Less abundant (20 %), Abundant (20 %) | - |
| Cupressus sp. | Lf | Gardens, Cultivated fields | Scarce (50 %), Less abundant (25 %), Abundant (25 %) | - |
| <i>Cymbopogon citratus</i> (DC.) Stapf | Wp | Cultivated fields, Fallow, Swamp, Gardens | Scarce (40 %), Less abundant (35 %), Abundant (25 %) | - |
| <i>Eucalyptus globulus</i> subsp. <i>maidenii</i> (F. Muell.) | Lf, Ec | Artificial afforestation, Fallow | Scarce (50 %), Less abundant (35 %), Abundant (15 %) | - |
| <i>Euphorbia grantii</i> Oliv. | Lf, Latex | Cultivated fields, Gardens | Scarce (65 %), Less abundant (30 %), Abundant (5 %) | - |

| Species | Part used | Habitat | Perceptions of respondents on Abundance according to the survey (% of respondents) | Literature for the plants perceived as scarce |
|---|-----------|---|--|---|
| <i>Gymnanthemum amygdalinum</i> (Delile) Sch. Bip. ex Walp | Lf | Cultivated fields, Gardens, Fallow | Scarce (60 %), Less abundant (25 %), Abundant (15 %) | - |
| <i>Mangifera indica</i> L. | Lf | Gardens, Fallow, Cultivated fields | Less abundant (35 %), Abundant (65 %) | - |
| <i>Markhamia lutea</i> (Benth.) K. Schum. | Lf | Semi-natural forest, Cultivated fields | Scarce (70 %), Less abundant (25 %), Abundant (5 %) | - |
| Cultivated | | | | |
| <i>Moringa oleifera</i> Lam. | Lf | Gardens, Cultivated fields, Fallow | Scarce (60 %), Less abundant (25 %), Abundant (15 %) | - |
| <i>Persea americana</i> Mill. | Lf | Fallow, Gardens, Cultivated fields | Less abundant (5 %), Abundant (95 %) | - |
| Plectranthus barbatus Andrews | Lf | Little disturbed forests, Cultivated fields, Fallow | Scarce (50 %), Less abundant (25 %), Abundant (25 %) | - |
| Plectranthus esculentus N.E.Br. | Lf | Savannahs | Scarce (80 %), Less abundant (15 %), Abundant (5 %) | - |
| <i>Psidium guajava</i> L. | Lf | Little disturbed forests, Cultivated fields, Gardens, Savannahs | Less abundant (10 %), Abundant (90 %) | - |
| <i>Senna didymobotrya</i> (Fresen.) H. S. Irwin & Barneby | Lf | Cultivated fields, Little disturbed forests, Fallow | Scarce (80 %), Less abundant (20 %) | - |
| <i>Solanecio mannii</i> (Hook.f.) C. Jeffrey | Lf | Cultivated fields, Fallow, Gardens | Scarce (40 %), Less abundant (25 %), Abundant (35 %) | - |
| <i>Tetradenia urticifolia</i> (Hochst.) Codd | Lf | Cultivated fields, Fallow, Gardens | Scarce (20 %), Less abundant (45 %), Abundant (35 %) | - |
| Zea mays L. | Lf | Cultivated fields, Swamp | Less abundant (5 %), Abundant (95 %) | - |

Table 4. Chemical structures of antimalarial and mosquito repellent compounds isolated from some of the plant species listed in Burundi

| Plant species | Structures | Effect | References |
|---------------------------------------|---|--------------|---|
| Artemisia annua L. | $ \begin{array}{c} \begin{array}{c} & H_{3}CO \\ & & H_{3}CO \\ & & H_{3}CO \\ & & H_{3}CO \\ & & H_{3}CO$ | Antimalarial | Artemisinin (1) (Kohler <i>et al.</i> 1997; Willcox 2009); Artemetin (2) , casticin (3) , chrysoplenetin (4) , chrysosplsrtol-D (5) , cirsilmeol (6) , eupatorin (7) (Cao <i>et al.</i> 2020; Gruessner <i>et al.</i> 2021, Zhou <i>et al.</i> 2021) |
| <i>Azadirachta indica</i> A. Juss. | | Antimalarial | Azadirachtin (8) (Pohlit <i>et al.</i> 2011). |
| <i>Bidens pilosa</i> L. | O $(10) R = H$ $(10) R = H$ $(11) R = Glucose$ $(11) R = Glucose$ $(12) R = Alpha-L-rhamnopyranosyl-(1-6)-beta-D-glucopyrose$ $(13) R = Beta-D-glucopyranose$ (14) | Antimalarial | Phenylheptatriyne (9), (R)- 1,2-dihydroxytrideca- 3,5,7,9,11-pentayne (10), 2- β -D-glycopyrasyloxy- 1-hydroxytrideca- 3,5,7,9,11-pentayne (11) ; flavonoids (12, 13), Phenylacetylene (14) (Nogueira and Lopes 2011; Tobinaga <i>et al.</i> 2009; Antoniana U Krettli <i>et al.</i> 2001). |
| <i>Cajanus cajan</i> (L.) Huth | (14) $(15) R_{1} = CH_{2}CHC(CH_{3}), R_{2} = H$ $(16) R_{1} = H, R_{2} = CH_{2}CHC(CH_{3})_{2}$ $HO \qquad (17)$ (17) $(16) R_{1} = H, R_{2} = CH_{2}CHC(CH_{3})_{2}$ $HO \qquad (17)$ (17) (19) | Antimalarial | Longistylin A (15) & C (16) , and betulinic acid (17) (Duker-Eshun <i>et al.</i> 2004). Cajachalcone 2',6'- dihydroxy-4-methoxy chalcone (18) ; 2,4- dimethoxy-4'-butoxy chalcone (19) ; Licochalcone (20) (Ajaiyeoba <i>et al.</i> 2013). |

| Plant species | Structures | Effect | References |
|--|--|---------------------|---|
| <i>Carica papaya</i> L. | (21) (21) (21) (21) (22) (22) (22) (22) | Antimalarial | (1S-11R-13S-14S-24R- 26S)-13,26-dimethyl-2,15- dioxa-12,25-dia-zatricyclo [22.2.2.211,14] triacontane- 3,16-dione ((þ)-carpaine (21); 6-(8-Methoxy-8- oxooctyl)-2- methylpiperidin-3-yl8-(5- hydroxy-6- methylpiperidin-2-yl) octanoate (22); 13,26- dimethyl-2,15-dioxa-12,25- diazatricyclo [22.2.2.2 11,14] triacontane-3,16- dione (23) (Julianti <i>et al.</i> 2014) |
| <i>Cinchona officinalis</i> L. | | Antimalarial | Quinine (24) (Cragg and Newman 2005) |
| <i>Citrus limon</i> (L.) Osbeck | HO HO (27) (25) (26) | Insect repellent | Limonene (25), α-terpineol (26) and myrcene (27) (Pohlit <i>et al.</i> 2011) |
| Cupressus sp. | (25) (28) (29) (25) (28) (29) (29) (31) (32) | Insect repellent | Limonene (25), α & β- pinene (28 & 29), 3-carene (30), carvacrol (31) and terpinen-4-ol (32) (Pohlit <i>et al.</i> 2011). |
| <i>Cymbopogon</i> <i>citratus</i> (DC.) Stapf | HO (27) (33) | Insect repellent | Myrcene (27) and geranial (33) (Pohlit <i>et al.</i> 2011). |



Azadirachta indica A. Juss.

Azadirachta indica A. Juss. (Meliaceae) is a species native to India and is known as Neem. It has become naturalized in Burundi where it primarily serves as an insecticidal plant. It is also one of the main insecticidal plants (also mosquito repellent) recommended to farmers by INADES Formation Burundi to farmers as alternatives to chemical pesticides, some of which are classified as dangerous and toxic (INADES Formation Burundi 2022). Additionally, it is used primarily as a mosquito repellent in many parts of the world. The species has been reported to repel the female mosquito (*Anopheles stephensi*), which is the main vector of malaria in urban India (Pohlit *et al.* 2011). *Azadirachta indica* is widely marketed around the world in the form of vegetable oil with insecticidal properties because of the main component (azadirachtin, **2**) of the leaves, flowers and fruits (Isman 2006). The molecule acts as an anti-appetizing, repellent and loathsome agent. It also induces growth retardation, inhibition of moulting and malformations, sterility in insects (by preventing egg-laying and interrupting sperm production in men). It is a very interesting insecticide with a much lower environmental impact than synthetic pesticides (Chaudhary *et al.* 2017). Finally, this oil derived from the seeds of this species is one of the patented essential vegetable oils used in mosquito repellent inventions (Pohlit *et al.* 2011).

Bidens pilosa L.

Bidens pilosa L. (Asteraceae) is commonly known as **icanda** in Burundi. It is used in both traditional human and veterinary medicine to treat a variety of pathologies, including microbial infections (Ngezahayo *et al.* 2015). In addition, phytochemical screenings for antimalarial agents were performed on several plant species (including *B. pilosa*) in Brazil and showed that the plant possessed significant antimalarial activity both *in vivo* and *in vitro* (Krettli *et al.* 2001). Similar studies have also been conducted by other groups of researchers, and all have confirmed this effect of the plant, by testing its extracts *in vitro* (Clarkson et al. 2004; Oliveira *et al.* 2004; Lacroix *et al.* 2011), and *in vivo* (Brandão *et al.* 1997; Andrade-neto *et al.* 2004; Krettli 2009). In addition, compounds with potent antimalarial effects (Table 4, structures **9-14**) have been isolated and identified in extracts of the species (Krettli *et al.* 2001; Tobinaga *et al.* 2009; Nogueira and Lopes 2011).

Cajanus cajan (L.) Huth

Cajanus cajan (L.) Huth (Fabaceae) is a perennial seed legume grown in tropical regions. In addition to its reported use as a febrifuge in traditional medicine (Duke and Martinez 1994), antimalarial activity of the leaf extracts of this plant has also been reported (Ajaiyeoba *et al.* 2005). Antiplasmodial agents have also been isolated and characterized from the plant material. Thus, two stilbenes (longistylin A, **15** and C, **16**), and betulinic acid (**17**), isolated from the extracts of the roots and leaves of the plant, were tested for their antimalarial effects, and showed moderately high in vitro activity against chloroquine-sensitive *Plasmodium falciparum* strain 3D7 (Duker-eshun *et*

al. 2004). Three charcones (**18-20**) were also isolated from *C. cajan* leaf extract and a test on a strain of *Plasmodium falciparum* (multi-resistant strain K1) gave very good results (Ajaiyeoba *et al.* 2013).

Carica papaya L.

Carica papaya L. (Caricaceae) is a well-known fruit tree in tropical regions. It is also known for its several medicinal uses (Krishna *et al.* 2008). It is used in particular against fever in traditional Burundian medicine (Ngezahayo *et al.* 2015), and its leaves are used against malaria (Tor-anyiin *et al.* 2003; Bertani *et al.* 2005; Ellena *et al.* 2012; Suleman *et al.* 2018). In 2018, Woon-Chien Teng and his collegues isolated one of the plant's active components. They discovered that it was an alkaloid (carpaine, **21**) with good activity against two strains of *Plasmodium falciparum* 3D7 and Dd2 (Teng *et al.* 2019). Furthermore, this alkaloid had already been characterized in the plant in 2014. The antiplasmodial test revealed that carpain had high antiplasmodial activity and low cytotoxicity when compared to two other alkaloids (**22, 23**) isolated from the same plant extract. A synergistic effect between *C. papaya* extract and artesunate has also been reported. Indeed, short-term co-administration of the methanolic extract of the plant and artesunate in mice infected with *Plasmodium berghei* showed an increase in antimalarial efficacy (Oraebosi and Good 2021).

Cinchona officinalis L.

Cinchona officinalis L. (Rubiaceae) is known in Burundi under the vernacular name **ikinini**, which means quinine (name of the active substance of the plant against malaria). It is a native of South America where species of the genus *Cinchona* (including *C. officinalis*) were known for their febrifuge properties. The first generation of antimalarial drugs used in the world was, quinine **(24)**, a quinoline alkaloid isolated from the bark of these plants for the first time by Pelletier and Caventou (French chemists and pharmacists) in 1820.

Citrus limon (L.) Osbeckn

Citrus limon (L) Osbeckn(Rutaceae) is an evergreen tree whose main raw material is the yellow fruit (edible), essential oil (insect repellent) and juice (most often used as a condiment) (Klimek-Szczykutowicz *et al.* 2020). The plant is cited in several patents on mosquito repellent inventions (Pohlit *et al.* 2011). The repellent properties of the plant against mosquitoes have been reported, particularly in *Anopheles stephensi* (main vector mosquito of malaria in urban areas in India), *Aedes aegypti* (main vector mosquito of dengue fever, Zika virus infection, chikungunya and yellow fever) and *Culex quinquefasciatus* (mosquito vector of various diseases including Nile fever, Saint Louis encephalitis, avian malaria) (Oshaghi *et al.* 2003; Amer and Mehlhorn 2006). Among the components of the essential oil of the plant (β -pinene, γ -terpinene, limonene (**25**), α -terpineol (**26**), myrcene (**27**), D-dihydrocarvone, only those bearing numbers have proven mosquito repellent and deterrent properties (Pohlit *et al.* 2011).

Cupressus sp.

Cupressus is a genus of trees in the Cupressaceae family with over 25 species. Essential oils from these species have been scientifically proven to repel a range of mosquitoes including *Anopheles stephensi, Aedes aegypti* and *Culex quinquefasciatus* (Pohlit *et al.* 2011). As with other essential oil plants mentioned in this work, species of this genus are cited in many patents on mosquito repellent inventions. Their chemical composition is very diverse; it contains in particular thujopsene, eudesmol, E-(+)- α -atlantone; α , $\beta \otimes \gamma$ -himachalenes; α - $\otimes \beta$ -cedrenes; limonene (26), β -phellandrene, $\alpha \otimes \beta$ -pinene (28 \otimes 29), 3-carene (30); p-methyl- Δ -3-tetrahydro \otimes p-methyl acetophenones; hinokitiol, carvacrol (31), terpinen-4-ol (32), sabinene (Burfield 2002). It should be noted that only molecules with numbers after their names (corresponding to the structures in Table 4) have proven to be anti-mosquito and deterrent (Pohlit *et al.* 2011).

Cymbopogon citratus (DC.) Stapf

Cymbopogon citratus (DC.) Stapf (Poaceae) is a perennial herb with large leafy clumps that grow 30-50 cm tall. It is a plant species native to Asia, but it is currently cultivated in all the intertropical regions (Troupin 1988). This plant species is known for its properties to repel mosquitoes such as *Aedes aegypti, Aedes atlanticus, Aedes mitchellae, Anopheles stephensi, Anopheles darlingi,* and *Culex quinquefasciatus*). The chemical composition of *C. citratus* essential oils used in patented mosquito repellent inventions includes geranial **(33)**, neral, myricene **(27)**. However, only geranial and myricene mosquito repellent properties (Pohlit *et al.* 2011).

Eucalyptus globulus subsp. maidenii (F. Muell.) Kirkp.

Eucalyptus globulus subsp. *maidenii* (F. Muell.) Kirkp. (Myrtaceae) is an introduced species in Burundi. It is used in particular against fever and cough in Burundian traditional medicine (Ngezahayo *et al.* 2015). During the COVID-

19 pandemic it was also very coveted and commercialized in this country: the decoction of its leaves was used as fumigants. This plant is used as both an antimalarial and a mosquito repellent. A large number of patents containing the genus *Eucalyptus* have already been registered for this last property (Pohlit *et al.* 2011). Additionally, it has been reported that essential oils of *Eucalyptus* spp. are widely used to repel mosquitoes and other insects and contain several mosquito repellents compounds such as 1,8-cineole (**34**), p-menthane-3,8-diol (**35**), α -pinene (**28**), p-cymene (**36**), γ -terpinene (**37**), citronellol (**38**) and α -terpineol (**26**) (Batish *et al.* 2008).

Gymnanthemum amygdalinum (Delile) Sch. Bip. ex Walp

Gymnanthemum amygdalinum (Delile) Sch. Bip. ex Walp (Asteraceae) is a synonym of *Vernonia amygdalina* Delile; it is used in the fight against insects harmful to humans (including mosquitoes) (Ahishakiye *et al.* 2022), and microbial infections (including malarial fever) in traditional Burundian medicine (Ngezahayo *et al.* 2015). Extracts of the species have already shown antimalarial activity *in vitro* (Muthaura *et al.* 2015) and *in vivo* (Abosi and Raseroka 2003; Okpe *et al.* 2016). Four sesquiterpene lactones (vernodalin, **39**; vernodalol, **40**; vemolide, **41**, hydroxyvemolide, **42**) have been isolated from the plant, and exhibited antimalarial activity *in vivo* in chempanzees (Ohigashi *et al.* 1994).

Conclusion

This study highlighted the use of more antimalarial plants than mosquito repellent plants in Burundi. Some plants species reported in this study were not reported in previous studies and require further investigation before they are recommended for use.

The study also showed that traditional knowledge is primarily held by older people, highlighting the importance of investing more in this sector to prevent the disappearance of traditional medicine knowledge. The parts used and the preparation modes are not limiting factors in plants conservation. The exploitation of these species is practically not a problem because most of them are found in degraded forests, artificial afforestation, fallow land, gardens and cultivated fields; but rarely in natural forests. However, conservation efforts are recommended for species that are not yet cultivated by the people, although they are scarce in some localities according to respondents.

Limitations

This study had some limitations: the number of respondents was not the same in each region due to the environment's accessibility and the study did not sufficiently document the availability of the species identified in the study area because the results were mostly based on respondent's information.

Declarations

List of abbreviations: F: Frequency; MINEAGRIE: Ministere de l'Environnement, de l'Agriculture et de l'Elevage (Ministry of the Environment, Agriculture and Livestock in Burundi).

Consent for publication: Not applicable.

Availability of data and materials: All the data are presented in figures, tables and appendix in the manuscript and are available with the corresponding author.

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Authors' contributions:

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Literature cited

Abdillah S, Tambunan RS, Farida Y, Sandhiutami NMD, Dewi RM. 2015. Phytochemical screening and antimalarial activity of some plants traditionally used in Indonesia. Asian Pacific Journal of Tropical Disease 5:454-457. doi: 10.1016/S2222-1808(15)60814-3.

Abe R, Ohtani K. 2013. An ethnobotanical study of medicinal plants and traditional therapies on Batan Island, the Philippines. Journal of Ethnopharmacology 145:554-565. doi: 10.1016/j.jep.2012.11.029.

Abosi AO, Raseroka BH. 2003. In vivo antimalarial activity of *Vernonia amygdalina*. British Journal of Biomedical Science 60:89-91. doi: 10.1080/09674845.2003.11783680.

Adesina J, Jose A, Rajashekar Y, Ileke K. 2016. *Persea americana* (MilL.) seed extracts: Potential herbal larvicide control measure against Anopheles gambiae Giles, 1902 (Diptera: Culicidae) Malaria vector. International Journal of Mosquito Research 3:14-17.

Adinew GM. 2014. Antimalarial activity of methanolic extract of Phytolacca dodecandra leaves against *Plasmodium berghei* infected Swiss albino mice. International Journal of Pharmacology and Clinical Sciences 3:39-45.

Adomou AC. 2005. Vegetation patterns and environmental gradients in Benin: Implications for biogeography and conservation. [Wageningen]: Wageningen University. https://edepot.wur.nl/121707.

Afolayan FID, Oladokun A, Fasoranti E. 2020. Comparative in vivo antiplasmodial activities of different extracts of *Lawsonia inermis, Tithonia diversifolia* and *Nauclea latifolia* against *Plasmodium berghei*. African Journal of Biological Sciences 2:9-17. doi: 10.33472/AFJBS.2.1.2020.9-17

Ahishakiye R, Nkengurutse J, Irampagarikiye R, Ndayizeye G, Bukuru A, Vyizigiro T, Masharabu T. 2022. Ethnobotanical study of pesticidal plants against human harmful insects in Central Burundi. East African Journal of Science, Technology and Innovation 3:1-15.

Ajaiyeoba EO, Bolaji OM, Akinboye DO, Falade CO, Gbotosho GO, Ashidi JS, Okpako LC, Oduola OO, Falade MO, Itiola OA. 2005. In vitro anti-plasmodial and cytotoxic activities of plants used as antimalarial agents in the southwest Nigerian ethnomedicine. Journal of Natural Remedies 5:1-6.

Ajaiyeoba EO, Ogbole OO, Abiodun OO, Ashidi JS, Houghton PJ, Wright CW. 2013. Cajachalcone: An antimalarial compound from *Cajanus cajan* leaf extract. Journal of Parasitology Research 2013:1-5. doi: 10.1155/2013/703781.

Akhilraj A.R., Rukmini S., Amalraj A.R. 2021. Scarcity of Medicinal Plants: A threat to Ayurveda-an overview. International Journal of Current Medical and Pharmaceutical Research 7:5992-5998.

Akpuaka A, Ekwenchi MM, Dashak DA, Dildar A. 2013. Isolation and characterization of an oxime from the n-Hexane extract of *Azadirachta*. New York Science Journal 6:86-90.

Alesaeidi S, Sepide M. 2016. A Systematic review of Anti-Malarial Properties, Immunosuppressive Properties, Anti-Inflammatory Properties, and Anti-Cancer Properties of *Artemisia annua*. Electronic Physician 8: 3150-55. doi: 10.19082/3150.

Amer A, Mehlhorn H. 2006. Repellency effect of forty-one essential oils against *Aedes, Anopheles,* and *Culex* mosquitoes. Parasitology Research 99:478-490. doi: 10.1007/s00436-006-0184-1.

Amir HM, Grace OM, Wabuyele E, Manoko MLK. 2019. Ethnobotany of *Aloe* L. (Asphodelaceae) in Tanzania. South African Journal of Botany 122:330-335. doi: 10.1016/j.sajb.2019.01.038.

Amuka O, Okemo PO, Alex K, Mbugua PK. 2014. Ethnobotanical Survey of Selected Medicinal Plants used by Ogiek Communities in Kenya against Microbial Infections. Ethnobotany Research and Applications 12:627-641. https://ethnobotanyjournal.org/index.php/era/article/view/805.

Ancolio C, Azas N, Mahiou V, Ollivier E, Giorgio DC, Keita A, Timon-David P, Balansard G. 2002. Antimalarial activity of extracts and alkaloids isolated from six plants used in traditional medicine in Mali and Sao Tome. Phytotherapy Research 16:646-649. doi: 10.1002/ptr.1025.

Andrade-neto VF, Brandão MGL, Oliveira FQ, Casali VWD, Njaine B, Zalis MG, Oliveira LA, Krettli AU. 2004. Antimalarial Activity of *Bidens pilosa* L. (Asteraceae) Ethanol Extracts From Wild Plants Collected in Various Localities or Plants Cultivated in Humus Soil. Phytotherapy Research 18:634-639. doi: 10.1002/ptr.1510.

Ankrah NA, Nyarko AK, Addo PGA, Ofosuhene M, Dzokoto C, Marley E, Addae MM, Ekuban FA. 2003. Evaluation of efficacy and safety of a herbal medicine used for the treatment of malaria. Phytotherapy Research 17:697-701. doi: 10.1002/ptr.1196.

Bah S, Jäger AK, Adsersen A, Diallo D, Paulsen BS. 2007. Antiplasmodial and GABAA-benzodiazepine receptor binding activities of five plants used in traditional medicine in Mali, West Africa. Journal of Ethnopharmacology 110:451-457. doi: 10.1016/j.jep.2006.10.019.

Bahekar S, Kale R. 2013. Herbal Plants Used For the Treatment of Malaria- A Literature Review. Journal of Pharmacognosy and Phytochemistry 1:141-146.

Batish DR, Pal H, Kumar R, Kaur S. 2008. *Eucalyptus* essential oil as a natural pesticide. Forest Ecology and Management 256:2166-2174. doi: 10.1016/j.foreco.2008.08.008.

Bertani S, Bourdy G, Landau I, Robinson JC, Esterre P, Deharo E. 2005. Evaluation of French Guiana traditional antimalarial remedies. Journal of Ethnopharmacology 98:45-54. doi: 10.1016/j.jep.2004.12.020.

Betti JL, Iponga DM, Yongo OD, Mbomio DO, Yobo CM, Ngoy A. 2013. Ethnobotanical study of medicinal plants of the Ipassa- Makokou Biosphere Reserve, Gabon: Plants used for treating malaria. Journal of Medicinal Plants Research 7:2289-2292. doi: 10.5897/jmpr11.1754.

Bickii J, Tchouya GRF, Tchouankeu JC, Tsamo E. 2007. Antimalarial Activity in Crude Extracts of Some Cameroonian Medicinal Plants. African Journal of Traditional, Complementary and Alternative Medicines 4:107-111. doi: 10.4324/9781003232506-9.

Bihonegn T, Giday M, Yimer G, Animut A, Sisay M. 2019. Antimalarial activity of hydromethanolic extract and its solvent fractions of *Vernonia amygdalina* leaves in mice infected with Plasmodium berghei. Sage Open Medicine 7:1-10. doi: 10.1177/2050312119849766.

Bla K, Trebissou J, Bidie A, Assi Y, Zihiri-Guede N, Djaman A. 2015. Étude ethnopharmacologique des plantes antipaludiques utilisées chez les Baoulé- N'Gban de Toumodi dans le Centre de la Côte d'Ivoire. Journal of Applied Biosciences 85:7775. doi: 10.4314/jab.v85i1.4.

Bodeker G, Burford G, Chamberlain, Joanne Chamberlain JR, Bhat KKS. 2001. The underexploited medicinal potential of *Azadirachta indica* A. Juss. (Meliaceae) and *Acacia nilotica* (L.) Willd. ex Del. (Leguminosae) in sub-Saharan Africa: a case for a review of priorities. International Forestry Review 3:285-298.

Boniface PK, Verma S, Shukla A, Cheema HS, Srivastava SK, Khan F, Darokar MP, Pal A. 2015. Bioactivity-guided isolation of antiplasmodial constituents from *Conyza sumatrensis* (Retz.) E.H. Walker. Parasitology International Journal 64:118-123. doi: 10.1016/j.parint.2014.10.010.

Bonkian L, Yerbanga R, Coulibaly MT, Lefèvre T, Sangaré I, Ouédraogo T, Traore O, Ouédraogo J, Guiguemde T, Dabiré K. 2017. Plants against Malaria and Mosquitoes in Sahel region of Burkina Faso : An Ethno-botanical survey. International Journal of Herbal Medicine 5:82-87.

Bonkian L, Yerbanga R, Sangare I, Koama B, Soma A, Cisse M, Bamba S, Valea I, Tinto H, Ouedraogo J. 2019. In vivo Antiplasmodial and Insecticidal Activities of *Citrus limon* (L.) Osbeck (Rutaceae), Leaves Extracts. Scholarena Journal of *Pharmacy and Pharmacology* 6:1-5.

Brandão MGL, Krettli A., Soares LSR, Nery CGC, Marinuzzi HC. 1997. Antimalarial activity of extracts and fractions from *Bidens pilosa* and other *Bidens* species (Asteraceae) correlated with the presence of acetylene and flavonoid compounds. Journal of Ethnopharmacology 57:131-138. doi: 10.1016/S0378-8741(97)00060-3.

Burfield T. 2002. Cedarwood oils. Part, 1. 1:14-15.

Cao R, Hu H, Li Y, Wang X, Xu M, Liu J, Zhang H, Yan Y, Zhao L, Li W, Zhang T, Xiao D, Guo X, Li Y, Yang J, Hu Z, Wang M, Zhong W. 2020. Anti-SARS-CoV-2 Potential of Artemisinins In Vitro. American Chemical Society Infectious Disease *s* 6:2524-2531. doi:10.1021/acsinfecdis.0c00522.

Chaudhary S, Kanwar RK, Sehgal A, Cahill DM. 2017. Progress on Azadirachta indica Based Biopesticides in Replacing Synthetic Toxic Pesticides. Frontiers in Plant Science 8:1-13. doi: 10.3389/fpls.2017.00610.

Cheikhyoussef A, Shapi M, Matengu K, Ashekele HM. 2011. Ethnobotanical study of indigenous knowledge on medicinal plant use by traditional healers in Oshikoto region, Namibia. Journal of Ethnobiology and Ethnomedicine

7:10. doi: 10.1186/1746-4269-7-10.

Chiribagula V B, Amuri S B, Philippe ON, Byanga J K, Pierre D, Simbi JB L. 2020. Ethnobotanical study of plants used as antimalarial in traditional medicine in Bagira in Eastern RD Congo. Journal of Pharmacognosy and Phytochemistry 9:1-14. doi: 10.22271/phyto.2020.v9.i4a.11661.

Clarkson C, Maharaj VJ, Crouch NR, Grace OM, Pillay P, Matsabisa MG, Bhagwandin N, Smith PJ, Folb PI. 2004. In vitro antiplasmodial activity of medicinal plants native to or naturalised in South Africa. Journal of Ethnopharmacology 92:177-191. doi: 10.1016/j.jep.2004.02.011.

Clement OA, Anthony AE, Félicien MK, Mercy GT, Hedmon O, Anke W, Casim UT, Patrick EO. 2020. A review for selecting medicinal plants commonly used for malaria in Uganda. African Journal of Pharmacy and Pharmacology 14:347-361. doi: 10.5897/ajpp2020.5182.

Collaboration Research Group for Qinghaosu. 1977. A new sesquiterpene lactone-Qinghaosu [in Chinese]. Kexue Tongbao, Chinese Sci Bull. 3:142.

Cragg GM, Newman DJ. 2005. Biodiversity: A continuing source of novel drug leads. Pure Applied Chemistry 77:7-24. doi: 10.1351/pac200577010007.

Cudjoe E, Donu D, Okonu RE, Amponsah JA, Amoah LE. 2020. The in Vitro Antiplasmodial Activities of Aqueous Extracts of Selected Ghanaian Herbal Plants. Journal of Parasitology Research 2020: 5041919. doi: 10.1155/2020/5041919.

Cysne DN, Fortes TS, Reis AS, de Paulo Ribeiro B, dos Santos Ferreira A, do Amaral FMM, Guerra RNM, Marinho CRF, Nicolete R, Nascimento FRF. 2016. Antimalarial potential of leaves of *Chenopodium ambrosioides* L. Journal of Parasitology Research 115:4327-4334. doi: 10.1007/s00436-016-5216-x.

Czechowski T, Rinaldi MA, Famodimu MT, Van Veelen M, Larson TR, Winzer T, Rathbone DA, Harvey D, Horrocks P, Graham IA. 2019. Flavonoid Versus Artemisinin Anti-malarial Activity in *Artemisia annua* Whole-Leaf Extracts. Frontiers in Plant Science 10:1-11. doi: 10.3389/fpls.2019.00984.

Delbanco AS, Burgess ND, Cuni-Sanchez A. 2017. Medicinal Plant Trade in Northern Kenya: Economic Importance, Uses, and Origin. Economic Botany 71:13-31. doi: 10.1007/s12231-017-9368-0.

Dénou A, Koudouvo K, Togola A, Haïdara M, Dembélé S., Ballo FN, Sanogo R, Diallo D, Gbeassor M. 2017. Savoir traditionnel sur les plantes antipaludiques à propriétés analgésiques, utilisées dans le district de Bamako (Mali). Journal of Applied Biosciences 112:10985. doi: 10.4314/jab.v112i1.3.

Deshpande PK, Gothalwal R, Pathak AK. 2014. Phytochemical analysis and evaluation of antimalarial activity of *Azadirachta indica*. The Pharma Innovation Journal 3:12-16.

Dibessa TT, Engidawork E, Nedi T, Teklehaymanot T. 2020. Antimalarial activity of the aqueous extract of the latex of *Aloe pirottae* Berger. (Aloaceae) against *Plasmodium berghei* in mice. Journal of Ethnopharmacology 255:112763. doi: 10.1016/j.jep.2020.112763.

Dike IP, Obembe OO, Adebiyi FE. 2012. Ethnobotanical survey for potential anti-malarial plants in south-western Nigeria. Journal of Ethnopharmacology 144:618-626. doi: 10.1016/j.jep.2012.10.002.

Duke JA, Martinez RV. 1994. Amazonian Ethnobotanical Dictionary. CRC Press: Boca Raton, PL. :213.

Duker-eshun G, Jaroszewski JW, Asomaning WA, Oppong-boachie F, Christensen SB. 2004. Antiplasmodial Constituents of *Cajanus cajan*. Phytotherapy Research 18:128-130. doi: 10.1002/ptr.1375.

Dwivedi MK, Shyam BS, Shukla R, Sharma NK, Singh PK. 2020. GIS Mapping of Antimalarial Plants Based on Traditional Knowledge in Pushparajgarh Division, District Anuppur, Madhya Pradesh, India. Journal of Herbs, Spices and Medicinal Plants 26:356-378. doi: 10.1080/10496475.2020.1747583.

El-Emam MAW, Mahmoud SS, Bayaumy FEZA. 2015. Potential role of mefloquine (antimalarial drug) and methanol extract of *Chenopodium ambrosioides* and *Sesbania sesban* in mice infected with *Schistosoma mansoni*. Asian Pacific Journal of Tropical Disease 5:608-613. doi: 10.1016/S2222-1808(15)60898-2.

Elfawal MA, Towler MJ, Reich NG, Golenbock D, Weathers PJ, Rich SM. 2012. Dried Whole Plant *Artemisia annua* as an Antimalarial Therapy. PLoS One 7:1-7. doi: 10.1371/journal.pone.0052746.

Ellena R, Quave CL, Pieroni A. 2012. Comparative Medical Ethnobotany of the Senegalese Community Living in Turin (Northwestern Italy) and in Adeane (Southern Senegal). Evidence-Based Complementary and Alternative Medicine 2012:1-30. doi: 10.1155/2012/604363.

Falisse JB, Masino S, Ngenzebuhoro R. 2018. Indigenous medicine and biomedical health care in fragile settings: Insights from Burundi. Health Policy and Planning 33:483-493. doi: 10.1093/heapol/czy002.

Feng X, Cao S, Qiu F, Zhang B. 2020. Traditional application and modern pharmacological research of *Artemisia annua* L. Pharmacology and Therapeutics 216:107650. doi: 10.1016/j.pharmthera.2020.107650.

Frausin G, Ari DFH, Lima RBS, Kinupp VF, Ming LC, Pohlit AM, Milliken W. 2015. An ethnobotanical study of antimalarial plants among indigenous people on the upper Negro River in the Brazilian Amazon. Journal of Ethnopharmacology. 174:238-252. doi: 10.1016/j.jep.2015.07.033.

Geremedhin G, Bisrat D, Asres K. 2014. Isolation, characterization and in vivo antimalarial evaluation of anthrones from the leaf latex of *Aloe percrassa* Todaro. Journal of Natural Remedies 14:119-125. doi: 10.18311/jnr/2014/72.

Gou Y, Li Z, Fan R, Guo C, Wang L, Sun H, Li J, Zhou C, Wang C, Wang Y. 2020. Ethnobotanical survey and evaluation of traditional mosquito repellent plants of Dai people in Xishuangbanna, Yunnan Province, China. Journal of Ethnopharmacology. 262:113124. doi: 10.1016/j.jep.2020.113124.

Gruessner BM, Weathers PJ. 2021. In vitro analyses of *Artemisia* extracts on *Plasmodium falciparum* suggest a complex antimalarial effect. PLoS ONE 16: e0240874. doi:10.1371/journal.pone.0240874.

Gurmu AE, Kisi T, Shibru H, Graz B, Willcox M. 2018. Treatments used for malaria in young Ethiopian children: A retrospective study 11 Medical and Health Sciences 1103 Clinical Sciences. Malaria Journal 17:1-8. doi: 10.1186/s12936-018-2605-x.

Hamel T, Sadou S, Seridi R. 2018. Pratique traditionnelle d'utilisation des plantes médicinales dans la population de la péninsule de l'Edough (nord-est algérien). Ethnopharmacologia 59:75-81.

Hemingway J, Shretta R, Wells TNC, Bell D, Djimdé AA, Achee N, Qi G. 2016. Tools and Strategies for Malaria Control and Elimination: What Do We Need to Achieve a Grand Convergence in Malaria? PLoS Biology 14:1-14. doi: 10.1371/journal.pbio.1002380.

Idowu OA, Soniran OT, Ajana O, Aworinde DO. 2010. Ethnobotanical survey of antimalarial plants used in Ogun State, Southwest Nigeria. African Journal of Pharmacy and Pharmacology 4:55-60.

INADES Formation Burundi. 2022. Rapport annuel 2021. https://www.inadesformation.net/.

Isman MB. 2006. Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated world. Annual Review of Entomology 51:45-66. doi: 10.1146/annurev.ento.51.110104.151146.

Iyamah PC, Idu M. 2015. Ethnomedicinal survey of plants used in the treatment of malaria in Southern Nigeria. Journal of Ethnopharmacology 173:287-302. doi: 10.1016/j.jep.2015.07.008.

Julianti T, Mieri M De, Zimmermann S, Ebrahimi SN. 2014. HPLC-based activity pro fi ling for antiplasmodial activity in the traditional Indonesian medicinal plant Carica papaya L. Journal of Ethnopharmacology 8:426-434. doi: 10.1016/j.jep.2014.05.050.

Kalonda EM, Mbayo MK, Muhume SK, Kasereka M, Mulamba J., Manya H., Lumbu JB., Misakabu F., Kasali M., Mpiana PT. 2014. Ethnopharmacological survey of plants used against malaria in Lubumbashi city (D.R. Congo). Journal of advanced Botany and Zoology 1:1-11. doi: 10.15297/jabz.v1i2.02.

Karle JM, Bhattacharjee AK. 1999. Stereoelectronic features of the cinchona alkaloids determine their differential antimalarial activity. Bioorganic and Medicinal Chemistry 7:1769-1774. doi: 10.1016/S0968-0896(99)00120-0.

Kasali F. 2014. Ethnopharmacological Survey of Medicinal Plants Used against Malaria in Bukavu City (D.R. Congo). European Journal of Medicinal Plants 4:29-44. doi: 10.9734/ejmp/2014/5766.

Katuura E, Waako P, Tabuti JRS, Bukenya-ziraba R, Ogwal-okeng J. 2007. Antiplasmodial activity of extracts of selected medicinal plants used by local communities in western Uganda for treatment of malaria. African Journal of Ecology 45:94-98. doi: 10.1111/j.1365-2028.2007.00864.x.

Kaushik NK, Bagavan A, Rahuman AA, Zahir AA, Kamaraj C, Elango G, Jayaseelan C, Kirthi A V., Santhoshkumar T,

Marimuthu S. 2015. Evaluation of antiplasmodial activity of medicinal plants from North Indian Buchpora and South Indian Eastern Ghats. Malaria Journal 14:1-8. doi: 10.1186/s12936-015-0564-z.

Kavana PY, Sangeda AZ, Mtengeti EJ, Mahonge C, Bukombe J, Fyumagwa R, Nindi S. 2019. Herbaceous plant species diversity in communal agro-pastoral and conservation areas in western Serengeti, Tanzania. Tropical Grasslands-Forrajes Tropicales 7:502-518. doi: 10.17138/TGFT(7)502-518.

Kiraithe MN, Nguta JM, Mbaria JM, Kiama SG. 2016. Evaluation of the use of *Ocimum suave* Willd. (Lamiaceae), *Plectranthus barbatus* Andrews (Lamiaceae) and *Zanthoxylum chalybeum* Engl. (Rutaceae) as antimalarial remedies in Kenyan folk medicine. Journal of Ethnopharmacology 178:266-271. doi: 10.1016/j.jep.2015.12.013.

Klimek-Szczykutowicz M, Szopa A, Ekiert H. 2020. *Citrus limon* (Lemon) Phenomenon—A Review of the Chemistry, Pharmacological Properties, Applications in the Modern Pharmaceutical, Food, and Cosmetics Industries, and Biotechnological Studies. Plants 9:1-24. doi: 10.3390/plants9010119.

Koch A, Tamez P, Pezzuto J, Soejarto D. 2005. Evaluation of plants used for antimalarial treatment by the Maasai of Kenya. Journal of Ethnopharmacology 101:95-99. doi: 10.1016/j.jep.2005.03.011.

Kohler M, Haerdi W, Christen P, Veuthey J. 1997. Extraction of artemisinin and artemisinic acid from *Artemisia annua* L. using supercritical carbon dioxide. Journal of Chromatography. 785:353-360. doi: 10.1016/S0021-9673(97)00403-2.

Kouadio B, Djeneb C, Yvette FNB, Yao K, Cynthia YY, Alain AS, Noël ZG. 2016. Étude ethnobotanique des plantes médicinales utilisées dans le Département de Transua, District du Zanzan (Côte d'Ivoire) Journal of Animal and Plant Sciences 27:4230-4250.

Koudouvo K, Karou DS, Kokou K, Essien K, Aklikokou K, Glitho IA, Simpore J, Sanogo R, De Souza C, Gbeassor M. 2011. An ethnobotanical study of antimalarial plants in Togo Maritime Region. Journal of Ethnopharmacology 134:183-190. doi: 10.1016/j.jep.2010.12.011.

Krettli AU, Andrade-neto VF, Brandão GL, Ferrari WMS. 2001. The Search for New Antimalarial Drugs from Plants Used to Treat Fever and Malaria or Plants Ramdomly Selected : a Review. Memórias do Instituto Oswaldo Cruz, Rio Janeiro 96:1033-1042. doi: 10.1590/S0074-02762001000800002.

Krettli AU. 2009. Antimalarial drug discovery: screening of Brazilian medicinal plants and purifi ed compounds. Expert Opinion on Drug Discovery 5:95-108. doi: 10.1517/17530050802678127.

Krishna KL, Paridhavi M, Patel JA. 2008. Review on nutritional, medicinal and pharmacological properties of Papaya (*Carica papaya* Linn.). Natural Product Radiance 7:364-373.

Kweka EJ, Mosha F, Lowassa A, Mahande AM, Kitau J, Matowo J, Mahande MJ, Massenga CP, Tenu F, Feston E. 2008. Ethnobotanical study of some of mosquito repellent plants in north-eastern Tanzania. Malaria Journal 7:1-9. doi: 10.1186/1475-2875-7-152.

Lacroix D, Prado S, Kamoga D, Kasenene J, Namukobe J, Krief S, Dumontet V, Mouray E, Bodo B, Brunois F. 2011. Antiplasmodial and cytotoxic activities of medicinal plants traditionally used in the village of Kiohima, Uganda. Journal of Ethnopharmacology 133:850-855. doi: 10.1016/j.jep.2010.11.013.

Lapenna S, Bergonzi M C, Vincieri F F, Bilia A R. 2008. Natural Product Communications Comparative Analysis of Antimalarial Principles in Artemisia annua L . Herbal Drugs from East Africa. Natural Product Communications 3:2003-2006. https://journals.sagepub.com/doi/pdf/10.1177/1934578X0800301213.

Latorre EC, Canavero A, Pochettino ML. 2018. Comparison of medicinal plant knowledge between rural and urban people living in the Biosphere Reserve Bioma Pampa Quebradas del Norte, Uruguay: an opportunity for biocultural conservation. Ethnobiology and Conservation 7:1-6. doi: 10.15451/ec2018.

Latreche M, Sadoudi Z. 2017. Etude Ethnobotanique et Caractéristique Phytochimique des Plantes Médicinales à effet Antimicrobien. Université Akli Mohand Oulhadj, Bouira, Algérie.

Lebrun J.P., Stork L. 1991. Enumération des plantes à fleurs d'Afrique tropicale. Editions des conservatoire et Jardin botaniques de Genève, Genève.

Liu W, Liu Y. 2015. Youyou Tu: significance of winning the 2015 Nobel Prize in Physiology or Medicine. Cardiovascular Diagnosis and Therapy 6:1-2. doi:10.3978/j.issn.2223-3652.2015.12.11.

Lusakibanza M, Mesia G, Tona G, Karemere S, Lukuka A, Tits M, Angenot L, Frédérich M. 2010. In vitro and in vivo antimalarial and cytotoxic activity of five plants used in congolese traditional medicine. Journal of Ethnopharmacology 129:398–402. doi:10.1016/j.jep.2010.04.007.

Lusakibanza M, Mesia G, Tona G, Karemere S, Lukuka A, Tits M, Angenot L, Frédérich M. 2010. In vitro and in vivo antimalarial and cytotoxic activity of five plants used in congolese traditional medicine. Journal of Ethnopharmacology 129:398-402. doi: 10.1016/j.jep.2010.04.007.

Lutgen P, Chan D, Sadr A. 2018. Effects of silver diammine fluoride on bond strength of adhesives to sound dentin. Dental Materials Journal 37:1003-1009. doi: 10.4012/dmj.2017-401.

Malann YD, Matur BM, Deme GG, Mailafia S. 2015. Antiplasmodial activity of extracts and fractions of *Casuarina* equistifolia against *Plasmodium berghei* in Mice. British Journal of Science 12:62-69.

Manouan NJ, Guessan N, Tiembré I. 2014. Identification des acteurs de la médecine traditionnelle en Côte d'Ivoire : cas du District Autonome d'Abidjan. Abidjan, Côte d'Ivoire.

Manya MH, Keymeulen F, Ngezahayo J, Bakari AS, Kalonda ME, Kahumba BJ, Duez P, Stévigny C, Lumbu SJB. 2020. Antimalarial herbal remedies of Bukavu and Uvira areas in DR Congo: An ethnobotanical survey. Journal of Ethnopharmacology 249:112422. doi: 10.1016/j.jep.2019.112422.

Maroyi A, Asteraceae K, Moore S, Moore VS, O V, Moore VS, Hoffm VO, Wild D, Moore VS, Moore S. 2020. *Baccharoides lasiopus*: Review of its Medicinal Phytochemistry and Pharmacological Properties. Journal of Pharmacy and Nutrition Sciences 10:205-212.

Masharabu T, Bigendako MJ, Lejoly J, Nkengurutse J, Noret N, Bizuru E, Bogaert J. 2010. Etude analytique de la flore et de la végétation du Parc National de la Ruvubu, Burundi. International Journal of Biological and Chemical Sciences 4:834-856.

Mavundza EJ, Maharaj R, Finnie JF, Kabera G, Van Staden J. 2011. An ethnobotanical survey of mosquito repellent plants in uMkhanyakude district, KwaZulu-Natal province, South Africa. Journal of Ethnopharmacology 137:1516-1520. doi: 10.1016/j.jep.2011.08.040.

Melariri P, Campbell W, Etusim P, Smith P. 2011. In vitro and in vivo antiplasmodial activities of extracts of *Cymbopogon citratus* Staph and *Vernonia amygdalina* Delile leaves. Journal of Natural Products 4:164-172.

Mesfin A, Giday M, Animut A, Teklehaymanot T. 2012. Ethnobotanical study of antimalarial plants in Shinile District, Somali Region, Ethiopia, and in vivo evaluation of selected ones against *Plasmodium berghei*. Journal of Ethnopharmacology 139:221-227. doi: 10.1016/j.jep.2011.11.006.

Michayewicz N. 2013. L'Aloe vera, plante médicinale traditionnellement et largement utilisée depuis des millénaires, aux nombreuses propriétés thérapeutiques. Plante miracle? Sciences pharmaceutiques. https://hal.univ-lorraine.fr/hal-01731937.

MINEAGRIE. 2014. Etat des Ressources Génétiques Forestières dans le Monde: Rapport national. Bujumbura, Burundi.

Mpondo EM, Ngene JP, Som LM, Loe GE, Céleste P, Boumsong NGO, Yinyang J, Dibong SD. 2017. Connaissances et usages traditionnels des plantes médicinales du département du haut Nyong. Journal of Applied Biosciences 113:11229-11245. doi: 10.4314/jab.v113i1.12.

Muganga R, Angenot L, Tits M, Frédérich M. 2010a. Antiplasmodial and cytotoxic activities of Rwandan medicinal plants used in the treatment of malaria. Journal of Ethnopharmacology 128:52-57. doi: 10.1016/j.jep.2009.12.023.

Mukungu N, Abuga K, Okalebo F, Ingwela R, Mwangi J. 2016. Medicinal plants used for management of malaria among the Luhya community of Kakamega East sub-County, Kenya. Journal of Ethnopharmacology 194:98-107. doi: 10.1016/j.jep.2016.08.050.

Mulei JM, Otieno DF, Onkware AO. 2014. An ethnobotanical study of the swamp wetland vegetation of Uasin Gishu County, Kenya. Ethnobotany Research and Applications 12:315-324.

Muregi FW, Ishih A, Miyase T, Suzuki T, Kino H, Amano T, Mkoji GM, Terada M. 2007. Antimalarial activity of methanolic extracts from plants used in Kenyan ethnomedicine and their interactions with chloroquine (CQ) against a CQ-tolerant rodent parasite, in mice. Journal of Ethnopharmacology 111:190-195. doi: 10.1016/j.jep.2006.11.009.

Murugan K, Panneerselvam C, Samidoss CM, Madhiyazhagan P, Suresh U, Roni M, Chandramohan B, Subramaniam J, Dinesh D, Rajaganesh R. 2016. In vivo and in vitro effectiveness of Azadirachta indica-synthesized silver nanocrystals against *Plasmodium berghei* and *Plasmodium falciparum*, and their potential against malaria mosquitoes. Research in Veterinary Science 106:14-22.

Muthaura C, Keriko J, Mutai C, Yenesew A, Gathirwa J, Irungu B, Nyangacha R, Mungai G, Derese S. 2015. Antiplasmodial potential of traditional phytotherapy of some remedies used in treatment of malaria in Meru-Tharaka Nithi county of Kenya. Journal of Ethnopharmacology 175:315-323.

Nadia NAC, Cédric Y, Raoul SNS, Christian NO, Azizi MA, Diane GDC, Nkouayep VR, Jeanette Y, Gabriel TH, Mbida M. 2020. Antimalarial Activity of Ethyl Acetate Extract and Fraction of *Bidens pilosa* against *Plasmodium berghei* (ANKA). Journal of Parasitology Research 2020:1-8. doi: 10.1155/2020/8832724.

Namsa ND, Mandal M, Tangjang S. 2011. Anti-malarial herbal remedies of northeast India, Assam: An ethnobotanical survey. Journal of Ethnopharmacology 133:565-572. doi: 10.1016/j.jep.2010.10.036.

Ndayizeye G, Imani G, Nkengurutse J, Irampagarikiye R, Ndihokubwayo N, Niyongabo F, Cuni-Sanchez A. 2020. Ecosystem services from mountain forests: Local communities' views in Kibira National Park, Burundi. Ecosystem Services 45:101171. doi: 10.1016/j.ecoser.2020.101171.

Ndiaye I, Camara B, Ngom D. 2017. Diversité spécifique et usages ethnobotaniques des ligneux suivant un gradient pluviométrique Nord-Sud dans le bassin arachidier sénégalais. Journal of Applied Biosciences 113:11123-11137. doi: 10.4314/jab.v113i1.2.

Ng XY, Lua HK, Boo CM, Lim RCJ. 2021. Status and distribution of Dodonaea viscosa Jacq. Nature in Singapore 14: e2021039. doi: 10.26107/NIS-2021-0039.

Ngezahayo J, Havyarimana F, Hari L, Stévigny C, Duez P. 2015. Medicinal plants used by Burundian traditional healers for the treatment of microbial diseases. Journal of Ethnopharmacology 173:338-351. doi: 10.1016/j.jep.2015.07.028.

Njoroge GN, Bussmann RW. 2006. Diversity and utilization of antimalarial ethnophytotherapeutic remedies among the Kikuyus (Central Kenya). Journal of Ethnobiology and Ethnomedicine 2:1-7. doi: 10.1186/1746-4269-2-8.

Nogueira CR, Lopes LMX. 2011. Antiplasmodial Natural Products. Molecules 16:2146-2190. doi: 10.3390/molecules16032146.

Nondo RSO, Zofou D, Moshi MJ, Erasto P, Wanji S, Ngemenya MN, Titanji VPK, Kidukuli AW, Masimba PJ. 2015. Ethnobotanical survey and in vitro antiplasmodial activity of medicinal plants used to treat malaria in Kagera and Lindi regions, Tanzania. Journal of Medicinal Plants Research 9:179-192. doi: 10.5897/jmpr2014.5685.

Noronha M, Pawar V, Prajapati A, Subramanian RB. 2020. A literature review on traditional herbal medicines for malaria. South African Journal of Botany 28:292-303. doi: 10.1016/j.sajb.2019.11.017.

Nyambati GK, Lagat ZO, Maranga RO, Samuel M, Ozwara H. 2013. In vitro anti-plasmodial activity of *Rubia cordifolia, Harrizonia abyssinica, Leucas calostachys* and *Sanchus schweinfurthii* medicinal plants. Journal of Applied Pharmaceutical Science 3:57-62. doi: 10.7324/JAPS.2013.31210.

Nzigidahera B. 2012. Description du Burundi : Aspects physiques, Bujumbura. Burundi.

Nzuki BF. 2016. Recherches ethnobotaniques sur les plantes médicinales dans la Région de Mbanza- Ngungu, RDC. Belgique.

Odoh UE, Uzor PF, Eze CL, Akunne TC, Onyegbulam CM, Osadebe PO. 2018. Medicinal plants used by the people of Nsukka Local Government Area, south-eastern Nigeria for the treatment of malaria: An ethnobotanical survey. Journal of Ethnopharmacology 218:1-15. doi: 10.1016/j.jep.2018.02.034.

Odugbemi TO, Akinsulire OR, Aibinu IE, Fabeku PO. 2011. Medicinal plants useful for malaria therapy in Okeigbo, Ondo state, Southwest Nigeria. Plant Biology 4:191-198. doi: 10.1111/j.1438-8677.2011.00470.x.

Ohigashi H, Huffman MA, Izutsu D, Koshimizu K, Kawanaka M, Sugiyama H, Kirby GC, Warhurst DC, Allen D, Wright CW. 1994. Toward the chemical ecology of medicinal plant use in chimpanzees : the case of *Vernonia amygdalina*, a plant used by wild chimpanzees possibly for parasite-related diseases. Journal of Chemical Ecology 20:541-53.

Okello D, Kang Y. 2019. Exploring Antimalarial Herbal Plants across Communities in Uganda Based on Electronic Data. Evidence-Based Complementary and Alternative Medicine 2019:3057180. doi: 10.1155/2019/3057180.

Okokon JE, Antia BS, Mohanakrishnan D, Sahal D. 2017. Antimalarial and antiplasmodial activity of husk extract and fractions of *Zea mays*. Pharmaceutical Biology 55:1394-1400.

Okokon JE, Bassey U, Udobang JA, Bankehde HK. 2019. Antimalarial and Antipyretic Activities of Cornsilk Extract and Fractions of *Zea mays*. Discovery Phytomedicine 6:143-150.

Okpe O, Habila N, Ikwebe J, Upev VA, Okoduwa SIR, Isaac OT. 2016. Antimalarial Potential of *Carica papaya* and *Vernonia amygdalina* in Mice Infected with *Plasmodium berghei*. Journal of Tropical Medicine 2016:1-6. http://dx.doi.org/10.1155/2016/873897.

Oladeji OS, Oluyori AP, Bankole DT, Afolabi TY. 2020. Natural Products as Sources of Antimalarial Drugs: Ethnobotanical and Ethnopharmacological Studies. Scientifica 2020 :1-22. doi: 10.1155/2020/7076139.

Oliveira FQ, Andrade-neto V, Krettli AU, Brandão MGL. 2004. New evidences of antimalarial activity of *Bidens pilosa* roots extract correlated with polyacetylene and flavonoids. Journal of Ethnopharmacology 93:39-42. doi: 10.1016/j.jep.2004.03.026.

Olorunnisola OS, Adetutu A, Balogun EA, Afolayan AJ. 2013. Ethnobotanical survey of medicinal plants used in the treatment of malarial in Ogbomoso, Southwest Nigeria. Journal of Ethnopharmacology 150:71-78. doi: 10.1016/j.jep.2013.07.038.

Olusi TA, Ibukunoluwa MR, Dada EO. 2016. Nutritional quality assessment and antiplasmodial activity of *Cajanus cajan* (L.) Huth., *Crescentia cujete* L. and *Myrianthus preussii* Engl. from Akure, Southwestern Nigeria. International Journal of Phytomedicine 7:449-458.

Omara T. 2020. Antimalarial Plants Used across Kenyan Communities. Evidence-Based Complementary and Alternative Medicine 2020:4538602. doi: 10.1155/2020/4538602.

Oraebosi MI, Good GM. 2021. Original paper Carica papaya augments anti-malarial efficacy of artesunate in *Plasmodium berghei* parasitized mice. Annals of Parasitology 67:295-303. doi: 10.17420/ap6702.342.

Orwa C., Mutua A, Kindt R, Jamnadass R, Anthony S. 2009. Agroforestree Database:a tree reference and selection guide version 4.0. http://www.worldagroforestry.org/sites/treedbs/treedbs/treedbs/ases.asp.

Oshaghi M, Ghalandari R, Vatandoost H, Shayeghi M, Kamali-nejad M, Tourabi-Khaledi H, Abolhassani M, Hashemzadeh M. 2003. Repellent Effect of Extracts and Essential Oils of Citrus limon (Rutaceae) and *Melissa officinalis* (Labiatae) Against Main Malaria Vector, *Anopheles stephensi* (Diptera : Culicidae). Iran Journal of Public Health 32:47-52.

Othman AS, Lin J wen, Franke-Fayard BM, Kroeze H, van Pul FJA, Chevalley-Maurel S, Ramesar J, Marin-Mogollon C, Jore MM, Morin MJ. 2018. Expression of full-length *Plasmodium falciparum* P48/45 in *P. berghei* blood stages: A method to express and evaluate vaccine antigens. Molecular and Biochemical Parasitology 224:44-49. doi: 10.1016/j.molbiopara.2018.07.009.

Otieno NE, Analo C. 2012. Local indigenous knowledge about medicinal plants in and around Kakamega forest in western Kenya. F1000Research 1:1-40. doi: 10.12688/f1000research.1-40.v1.

Owuor BO, Ochanda JO, Kokwaro JO, Cheruiyot AC, Yeda RA, Okudo CA, Akala HM. 2012. In vitro antiplasmodial activity of selected Luo and Kuria medicinal plants. Journal of Ethnopharmacology. 144(3):779-781. doi: 10.1016/j.jep.2012.09.045. http://dx.doi.org/10.1016/j.jep.2012.09.045.

Oyeyemi IT, Akinseye KM, Adebayo SS, Oyetunji MT, Oyeyemi OT. 2019. Ethnobotanical survey of the plants used for the management of malaria in Ondo State, Nigeria. South African Journal of Botany 124:391-401. doi: 10.1016/j.sajb.2019.06.003.

Philip K, Elizabeth MM, Cheplogoi PK, Samuel KT. 2017. Ethnobotanical Survey of Antimalarial Medicinal Plants Used in Butebo County, Eastern Uganda. European Journal of Medicinal Plants 21:1-22. doi: 10.9734/EJMP/2017/35368.

Pohlit AM, Lopes NP, Gama RA, Tadei WP, Ferreira V, Neto DA. 2011. Patent Literature on Mosquito Repellent Inventions which Contain Plant Essential Oils - A Review. Planta Medica 77:598-617. doi: 10.1055/s-0030-1270723.

Priyadarshi A, Ram B. 2018. a Review on Pharmacognosy, Phytochemistry and Pharmacological Activity of *Carica papaya* (Linn.) Leaf. International Journal of Pharmaceutical Sciences and Research 9:4071-4078. doi: 10.13040/IJPSR.0975-8232.9(10).4071-78.

Pulice G, Pelaz S, Matías-Hernández L. 2016. Molecular farming in Artemisia annua, a promising approach to improve anti-malarial drug production. Frontiers in Plant Science 7:1-17. doi: 10.3389/fpls.2016.00329.

Reekmans M, Niyongere L. 1983. Lexique vernaculaire des plantes vasculaires du Burundi. Université du Burundi. Bujumbura, Burundi.

République du Burundi. 2018. Incidence du paludisme au Burundi. Bujumbura, Burundi

Rhattas M, Douira A, Zidane L. 2016. Étude ethnobotanique des plantes médicinales dans le Parc National de Talassemtane (Rif occidental du Maroc). Journal of Applied Biosciences 97:9187-9211. doi: 10.4314/jab.v97i1.5

Saad AM, Mohammed MMD, Ghareeb MA, Ahmed WS, Farid MA. 2017. Chemical composition and antimicrobial activity of the essential oil of the leaves of *Cupressus macrocarpa* Hartweg. ex Gordon. Journal of Applied Pharmaceutical Science 7:207-212. doi: 10.7324/JAPS.2017.70928.

Schmelzer GH, Gurib-Fakim A. 2008. Ressources végétales de l'Afrique tropicale 11 (1): Plantes médicinales. http://www.worldagroforestry.org/sites/treedbs/treedbabases.asp.

Selvam JP, Durai M. 2018. Study of medicinal plants used for the treatment of malaria in the Tiruchirappalli region. World Journal of Pharmaceutical Research 7:757-767. doi: 10.20959/wjpr201812-12632.

Seyfe S, Toma A, Esaiyas A, Debela E, Fikru A, Eyado A. 2017. Phytochemical screening and in vivo antimalarial activities of crude extracts of Lantana trifolia root and *Premna oligotricha* leaves in *Plasmodium berghei* infected mice. Journal of Medicinal Plants Research 11:763-769. doi: 10.5897/JMPR2017.6519.

Shah A, Rahim S. 2017. Ethnomedicinal uses of plants for the treatment of malaria in Soon Valley, Khushab, Pakistan. Journal of Ethnopharmacology 200:84-106. doi: 10.1016/j.jep.2017.02.005.

Sinzinkayo D. 2018. Etat des lieux de la malaria au Burundi. http://www.ub.edu.bi/wp-content/uploads/2018/05/EtatdeslieuxdelaMalariaauBurundi.pdf.

Suleman S, Tufa TB, Kebebe D, Belew S, Mekonnen Y, Gashe F, Musa S, Wynendaele E, Duchateau L, Spiegeleer B De. 2018. Treatment of Malaria and Related Symptoms Using Traditional Herbal Medicine in Ethiopia. Journal of Ethnopharmacology 213:262-279. doi: 10.1016/j.jep.2017.10.034.

Süntar I. 2020. Importance of ethnopharmacological studies in drug discovery: role of medicinal plants. Phytochemistry Reviews 19:1199-1209. doi: 10.1007/s11101-019-09629-9.

Sylla Y, Silue DK, Ouattara K, Kone MW. 2018. Etude ethnobotanique des plantes utilisées contre le paludisme par les tradithérapeutes et herboristes dans le district d'Abidjan (Côte d'Ivoire). International Journal of Biological and Chemical Sciences 12:1380. doi: 10.4314/ijbcs.v12i3.25.

Tefera BN, Kim YD. 2019. Ethnobotanical study of medicinal plants in the Hawassa Zuria District, Sidama zone, Southern Ethiopia. Journal of Ethnobiology and Ethnomedicine 15:13-26. doi: 10.1186/s13002-019-0302-7.

Teka T, Bisrat D, Yeshak MY, Asres K. 2016. Antimalarial activity of the chemical constituents of the leaf latex of *Aloe pulcherrima* Gilbert and Sebsebe. Molecules 21:1-10. doi: 10.3390/molecules21111415.

Teng W-C, Chan W, Suwanarusk R, Ong A, Ho H-K, Russell B, Rénia L, Koh H-L. 2019. In vitro Antimalarial Evaluations and Cytotoxicity Investigations of *Carica papaya* Leaves and Carpaine. Natural Product Communications 14:4-7. doi: 10.1177/1934578X1901400110.

Tobinaga S, Sharma MK, Aalbersberg WGL, Watanabe K, Iguchi K, Narui K, Sasatsu M, Waki S. 2009. Isolation and Identification of a Potent Antimalarial and Antibacterial Polyacetylene from *Bidens pilosa*. Planta Medica 75:624-628. doi: 10.1055/s-0029-1185377.

Tor-anyiin TA, Sha'ato R, Oluma HOA. 2003. Ethnobotanical Survey of Anti-Malarial Medicinal Plants Amongst the Tiv People of Nigeria Ethnobotanical Survey of Anti-Malarial Medicinal Plants Amongst the Tiv People of Nigeria. Journal of Herbs, Spices and Medicinal Plants 10:61-74. doi: 10.1300/J044v10n03_07.

Troupin G. 1982. Plantae africanae. IX (Acanthaceae, Melastomataceae, Sapotaceae). Bulletin Du Jardin Botanique

National de Belgique 52:463-465. doi: 10.2307/3667896.

Troupin G. 1983. Flora of Rwanda. Spermatophyta. Musée Royal d'Afrique Centrale l'Afrique (Belgium).V2.

Troupin G. 1985. Flora of Rwanda. Spermatophyta. Musée Royal d'Afrique Centrale l'Afrique (Belgium).V3.

Troupin G. 1988. Flora of Rwanda. Spermatophyta. Musee Royal d'Afrique Centrale l'Afrique (Belgium).V4.

Tugume P, Kakudidi EK, Buyinza M, Namaalwa J, Kamatenesi M, Mucunguzi P, Kalema J. 2016. Ethnobotanical survey of medicinal plant species used by communities around Mabira Central Forest Reserve, Uganda. Journal of Ethnobiology and Ethnomedicine 12:1-29. doi: 10.1186/s13002-015-0077-4.

Ved A, Arsi T, Prakash O, Gupta A. 2018. A review on phytochemistry and pharmacological activity of *Lantana camara* Linn. International Journal of Pharmaceutical Sciences and Research 9:37-43. doi: 10.13040/IJPSR.0975-8232.9(1).37-43.

Waiganjo B, Moriasi G, Onyancha J, Elias N, Muregi F. 2020. Antiplasmodial and Cytotoxic Activities of Extracts of Selected Medicinal Plants Used to Treat Malaria in Embu County, Kenya. Journal of Parasitology Research 2020. doi: 10.1155/2020/8871375.

WHO. 2018. World malaria report 2018. https://www.who.int/publications/i/item/9789241565653.

WHO. 2020. World malaria report 2020. https://www.who.int/publications/i/item/9789240015791.

Willcox M. 2009. Artemisia Species : From Traditional Medicines to Modern Antimalarials and Back Again. Journal of Alternative and Complementary Medicine 15:101-109. doi: 10.1089/acm.2008.0327.

Woerdenbag HJ, Lugt CB, Pras N. 1990. *Artemisia annua* L: a source of novel antimalarial drugs. Pharmaceutisch Weekblad Scientific Edition 12:169-181. doi: 10.1007/BF01980041.

Yetein MH, Houessou LG, Lougbégnon TO, Teka O, Tente B. 2013. Ethnobotanical study of medicinal plants used for the treatment of malaria in plateau of Allada, Benin (West Africa). Journal of Ethnopharmacology 146:154-163. doi: 10.1016/j.jep.2012.12.022.

Youmsi RDF, Fokou PVT, Menkem EZ, Bakarnga-Via I, Keumoe R, Nana V, Boyom FF. 2017. Ethnobotanical survey of medicinal plants used as insects repellents in six malaria endemic localities of Cameroon. Journal of Ethnobiology and Ethnomedicine 13:1-15. doi: 10.1186/s13002-017-0155-x.

Zhou Y, Gilmore K, Ramirez S, Settels E, Gammeltoft KA, Pham L V, Fahnøe U, Feng S, Ofersgaard A, Trimpert J, Bukh J, Osterrieder K, Gottwein J M, Seeberger PH. 2021. In vitro efficacy of artemisinin-based treatments against SARS-CoV-2. Scientific Report 11. doi:10.1038/s41598-021-93361-y.

Zirihi GN, Mambu L, Guédé-Guina F, Bodo B, Grellier P. 2005. In vitro antiplasmodial activity and cytotoxicity of 33 West African plants used for treatment of malaria. Journal of Ethnopharmacology 98:281-285. doi: 10.1016/j.jep.2005.01.004.